

1 MIB-derived odor management based upon hydraulic regulation
2 in small drinking water reservoirs: principle and application

3 Jinping Lu^{a,d}, Ming Su^{a,d,*}, Yuliang Su^b, Jiao Fang^a, Michael Burch^c, Tengxin Cao^{a,d}, Bin Wu^b,
4 Jianwei Yu^{a,d}, Min Yang^{a,d,*}

^a*State Key Laboratory of Environmental Aquatic Chemistry, Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences, P.O. Box 2871, Beijing, 100085,*

^b*Zhuhai Water Environment Holdings Group Ltd., Zhuhai, 519020,*

^c*School of Biological Sciences, The University of Adelaide, SA, 5005,*

^d*University of Chinese Academy of Sciences, Beijing, 100049,*

5 **Supplementary Material**

6 Figures and/or tables are provided below as the supplementary evidences to the main text.

*Corresponding author

Email addresses: mingsu@rcees.ac.cn (Ming Su), yangmin@rcees.ac.cn (Min Yang)

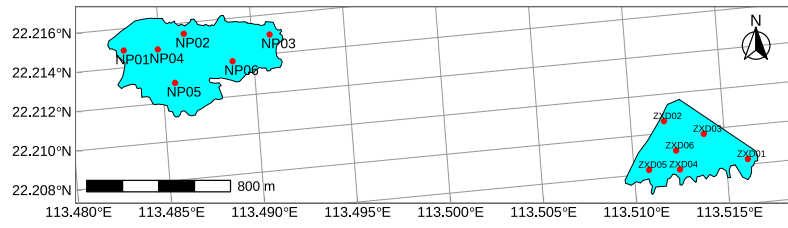


Fig. 1 The sampling sites in NP (A) and ZXD (B) Reservoirs

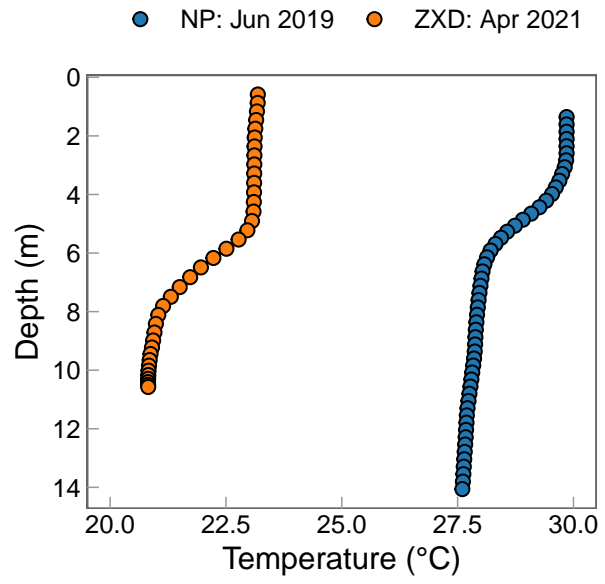


Fig. 2 Temperature profile of NP and ZXD Reservoirs

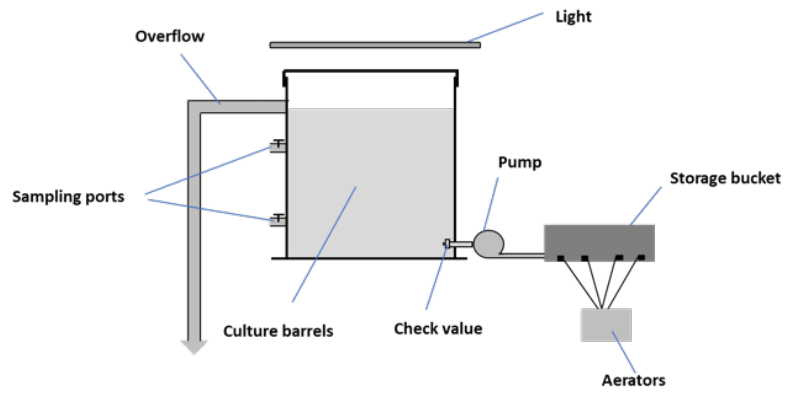


Fig. 3The schematic diagram of the experimental apparatus and the detailed design for the experiment

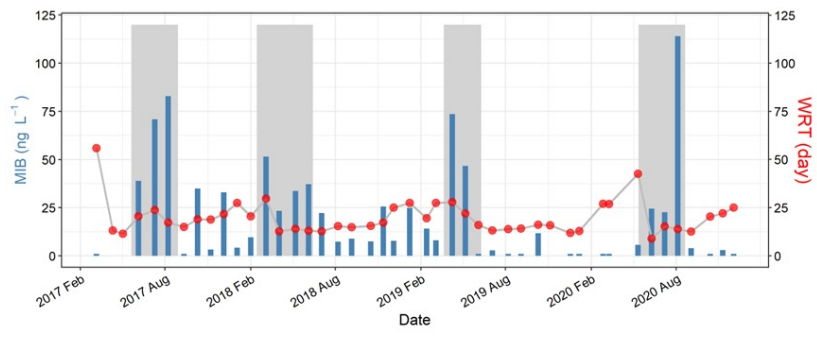


Fig. 4The monthly variation of water residence time and MIB from 2017 to 2020. (The MIB episodes are represented by grey areas)

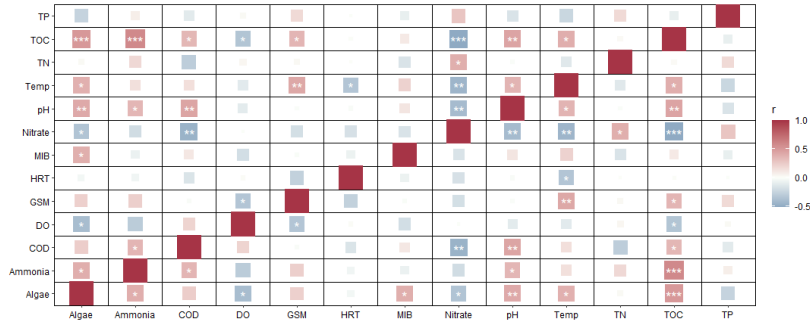


Fig. 5 The correlation analysis between MIB and some limnological parameters in NP Reservoir

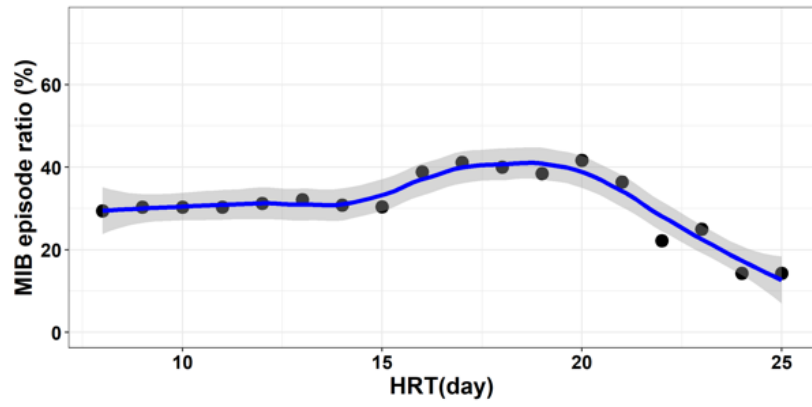


Fig. 6The relationship between MIB episode ration and HRT in NP Reservoir

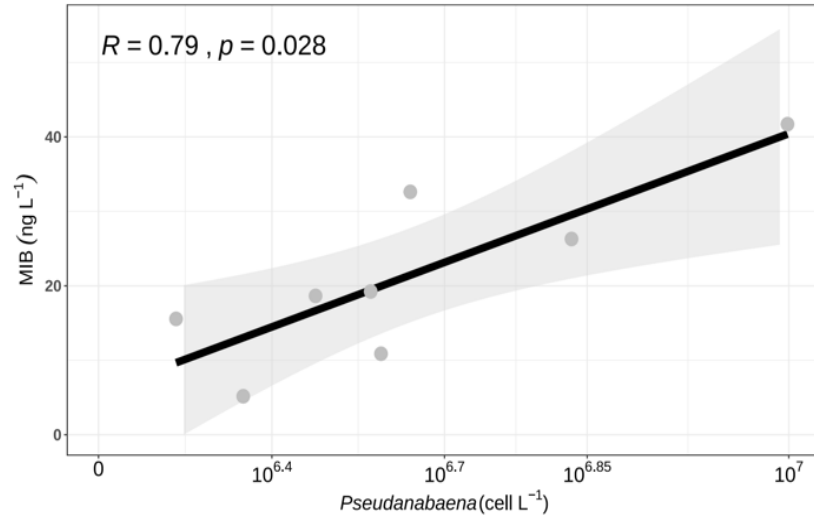


Fig. 7 The relationship between *Pseudanabaena* abundance and MIB concentration in ZXD Reservoir.

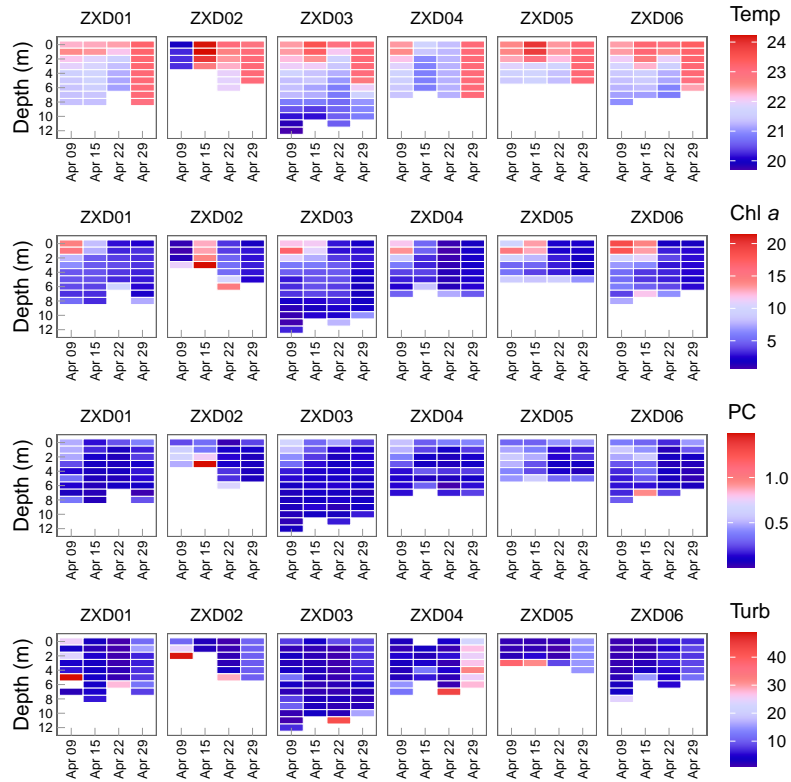


Fig. 8 The vertical variation of water quality parameters in ZXD Reservoir during the investigation period (April 2021) (A: Temperature (Temp); B : Chlorophyll *a* (Chl *a*); C : Phycocyanin (PC); D : Turbidity (Turb)).

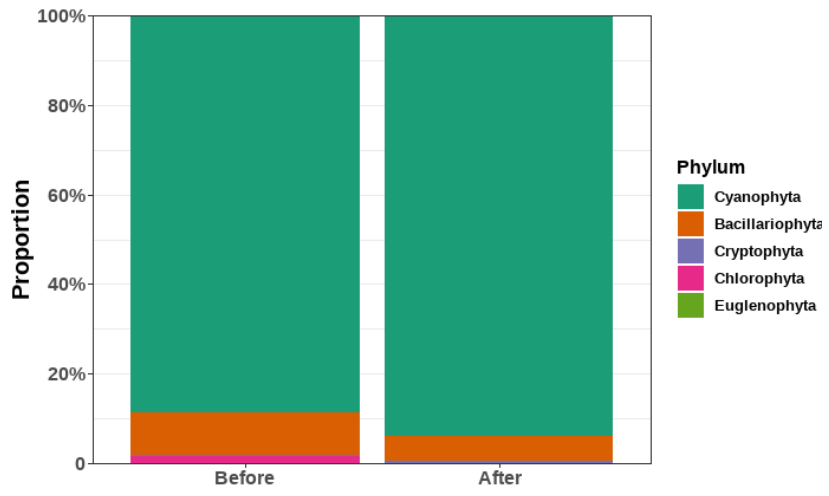


Fig. 9The phytoplankton phylum in ZXD Reservoir before and after HRT regulation

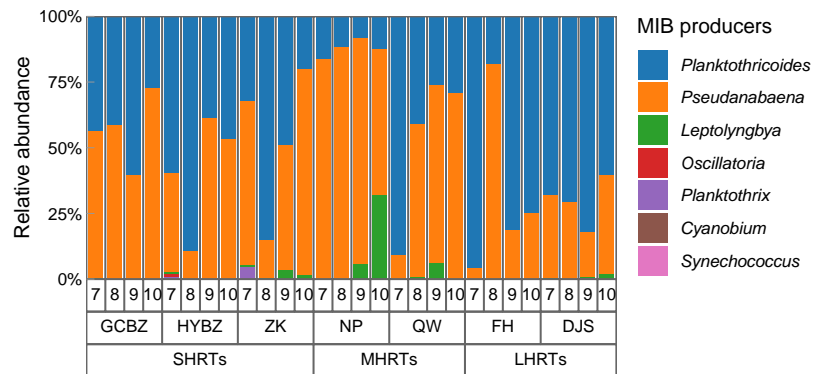


Fig. 10 The MIB-producing cyanobacteria genus in Zhuhai water sources

7 To identify the MIB-producing cyanobacteria genera in Zhuhai, seven water sources (GCBZ,
8 HYBZ, ..., FH, DJS) were select and water samples were collected from July to October, 2020 when
9 there were MIB episode. The universal primer for MIB gene (*mic*) (Suruzzaman et al., 2022) was
10 used to amplify samples genome and was Illumina MiSeq sequenced. The data was phylogenetic
11 classification assigned using Silva 128 database. The result declared that the MIB-producers in
12 Zhuhai was *Pseudanabaena* and *Planktothricoides* with the later more abundant.

Table 1 Sampling description

Reservoir	Sampling	Time	Sites	Layers	Frequency
NP	Routine	Mar., 2017~Dec., 2020	NP03	Surface (0.5 m)	monthly
NP	MIB episodes	Jul., 2017	NP01, NP02, ..., NP06	Surface (0.5 m)	daily
NP	MIB episodes	Apr., 2018	NP01, NP02, ..., NP06	Surface (0.5 m)	daily
ZXD	MIB episodes	Mar. ~ Apr. , 2021	ZXD01, ZXD02, ...,ZXD06	Surface (0.5 m)	daily
ZXD	MIB episodes	Apr. , 2021	ZXD01	Surface (0.5 m), middle (3 m), bottom (5~7 m)	weekly
12 ZXD	MIB episodes	Apr. , 2021	ZXD02	Surface (0.5 m), middle (2 m), bottom (3~5 m)	weekly
ZXD	MIB episodes	Apr. , 2021	ZXD03	Surface (0.5 m), middle (5 m), bottom (10~11 m)	weekly
ZXD	MIB episodes	Apr. , 2021	ZXD04	Surface (0.5 m), middle (3 m), bottom (6~7 m)	weekly

Reservoir	Sampling	Time	Sites	Layers	Frequency
ZXD	MIB episodes	Apr. , 2021	ZXD05	Surface (0.5 m), middle (2.5 m), bottom (5~6 m)	weekly
ZXD	MIB episodes	Apr. , 2021	ZXD06	Surface (0.5 m), middle (3.5 m), bottom (6~8 m)	weekly

Table 2 Light intensity for all sets was $54 \mu\text{mol m}^{-2} \text{s}^{-1}$, and culture temperature was $30 \text{ }^\circ\text{C}$; 10 samplings were performed for each set. The lab experiment design to investigate the effect of hydraulic residence time to *P. raciborskii* growth is shown in the table above.

ID	HRT (d)	Flowrate (L d ⁻¹)
I	2	4.5
II	5	1.8
III	10	0.9
IV	20	0.45
V	40	0.225
VI	80	0.113

13 The raw NP Reservoir was first collected and transported to laboratory, then filtered using
 14 glass fiber membrane ($0.7 \mu\text{m}$) to remove phytoplankton, the filtrate was further sterilized to
 15 remove microorganisms that may produce MIB, such as actinomycetes. Then fill the 9 L culture
 16 barrels with this sterilized solution and the rest served as the dilution solution was stored and
 17 aerated in the black storage bucket to minimize potential MIB accumulation. The enriched pure *P.*
 18 *raciborskii* was first filtered using polyester fiber membrane ($1.2 \mu\text{m}$, Millipore, USA) to remove
 19 excess nutrients from the original culture solution and washed 3 times with ultrapure water.
 20 Then *P. raciborskii* was cultured into the culture barrels at cell density of $1 \times 10^7 \text{ cell L}^{-1}$ with
 21 three replicates, the temperature and light intensity were $30 \text{ }^\circ\text{C}$ and $54 \mu\text{mol photon m}^{-2} \text{ s}^{-1}$. The
 22 residence time setting (2, 5, 10, 20, 40, 80 day) in different barrels was achieved by adjusting the
 23 flow rate (4.5, 1.8, 0.9, 0.45, 0.225, 0.113 L d⁻¹) of dilution solution pumped from the storage
 24 bucket. The 20 mL (10 mL for each sampling port and mixed into 20 mL) samples were taken
 25 every two days for odor compounds concentration and cell density quantification, the whole
 26 experiment was last for 19 days.

Table 3 The main water quality parameter in NP Reservoir (2017 ~ 2020, all values are expressed as mean values with standard deviations, variance analysis were performed to compare the differences (n = 71)).

Parameter	Spring	Summer	Fall	Winter	p-value
MIB _t (ng L ⁻¹)	27.1 ± 25.9	35.5 ± 36.1	9.27 ± 11.2	9.78 ± 11.2	p = 0.0334
GSM _t (ng L ⁻¹)	1.0 ± 0.0	2.9 ± 2.6	3.7 ± 2.9	1.9 ± 1.5	p = 0.0425
Temp. (°C)	22.5 ± 2.3	26.5 ± 1.3	25.1 ± 2.5	20.8 ± 3.7	p < 0.0001
TP (µg L ⁻¹)	41 ± 27	22 ± 13	36 ± 26	48 ± 44	p = 0.1863
TN (µg L ⁻¹)	1470 ± 167	1370 ± 500	1500 ± 455	1710 ± 357	p = 0.3051
TOC (mg L ⁻¹)	1.65 ± 0.16	1.76 ± 0.49	1.49 ± 0.29	1.40 ± 0.22	p = 0.0565
15 pH	8.5 ± 0.4	8.6 ± 0.5	8.1 ± 0.4	8.0 ± 0.4	p = 0.0023
NO ₃ -N (µg L ⁻¹)	1030 ± 389	872 ± 404	1150 ± 301	1410 ± 250	p = 0.0065
NH ₄ -N (µg L ⁻¹)	193 ± 77	159 ± 123	155 ± 123	94 ± 83	p = 0.2090
DO (mg L ⁻¹)	6.5 ± 1.5	6.6 ± 1.3	7.2 ± 1.4	7.3 ± 1.6	p = 0.4690
COD (mg L ⁻¹)	5.3 ± 1.4	6.1 ± 2.7	5.6 ± 1.9	5.2 ± 2.2	p = 0.7364
Phytoplankton (×10 ⁶ cell L ⁻¹)	39.7 ± 38.2	54.1 ± 48.8	13.6 ± 19.6	9.6 ± 15.4	p = 0.0084
HRT (d)	25.8 ± 13.8	15.3 ± 3.9	16.8 ± 2.8	21.8 ± 5.7	p = 0.0092

Table 4 The MIB-producing cyanobacteria genera identified by *mic* sequence in NP Reservoir

Phylum	Order	Genus	July	August
Cyanobacteria	Synechococcales	<i>Pseudanabaena</i>	83.6 %	88.3 %
Cyanobacteria	Oscillatoriales	<i>Planktothricoides</i>	16.4 %	11.7%

27 To identify the MIB-producing cyanobacteria genera in NP Reservoir, water samples were collected in July and August, 2020 when serious
 28 MIB episode occurred. The universal primer for MIB gene (*mic*) (Suruzzaman et al., 2022) was used to amplify samples genome and was
 29 Illumina MiSeq sequenced. The data was phylogenetic classification assigned using Silva 128 database. The result declared that the MIB-
 30 producers in NP Reservoir was *Pseudanabaena* and *Planktothricoides*.

Table 5 The differences of main limnological parameters characteristics between the periods with and without MIB episodes (MIB > 10 ng L⁻¹), all values are expressed as mean values with standard deviations, variance analysis were performed to compare the differences (n = 45).

Parameters	MIB episodes	No MIB episodes	<i>p</i> -value
MIB _t (ng L ⁻¹)	56.1 ± 26.2	8.3 ± 8.9	<i>p</i> < 0.0001
GSM _t (ng L ⁻¹)	3.2 ± 2.6	2.3 ± 2.2	<i>p</i> = 0.2721
Temp. (°C)	24.9 ± 3.2	23.2 ± 3.4	<i>p</i> = 0.1632
TP (µg L ⁻¹)	33 ± 26	38 ± 33	<i>p</i> = 0.6973
TOC (mg L ⁻¹)	1.68 ± 0.40	1.55 ± 0.33	<i>p</i> = 0.2857
TN (µg L ⁻¹)	1505 ± 442	1523 ± 403	<i>p</i> = 0.8980
¹⁷ pH	8.3 ± 0.5	8.2 ± 0.5	<i>p</i> = 0.5538
NO ₃ -N (µg L ⁻¹)	972 ± 277	1165 ± 408	<i>p</i> = 0.1570
NH ₄ -N (µg L ⁻¹)	151 ± 119	149 ± 107	<i>p</i> = 0.9675
DO (mg L ⁻¹)	6.1 ± 1.4	7.2 ± 1.4	<i>p</i> = 0.0257
COD (mg L ⁻¹)	5.6 ± 2.6	5.5 ± 1.8	<i>p</i> = 0.8614
Phytoplankton (×10 ⁶ cell L ⁻¹)	62.3 ± 52.1	18.1 ± 24.9	<i>p</i> = 0.0008
HRT (d)	20.2 ± 5.5	20.2 ± 9.8	<i>p</i> = 0.9987

Table 6 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus	P ₁₀	P ₅₀	P ₉₀
<i>Synechococcus</i>	0.17	0.72	3.06
<i>Limnothrix</i>	0.18	0.47	0.70
<i>Planktothrix</i>	0.16	0.44	0.71
<i>Aphanizomenon</i>	0.24	0.42	0.89
<i>Cylindrospermopsis</i>	0.14	0.36	0.74
<i>Plectonema</i>	0.11	0.34	2.06
<i>Microcystis</i>	0.10	0.33	0.67
<i>Planktothricoides</i>	0.17	0.33	1.18
<i>Microcoleus</i>	0.17	0.29	1.25
<i>Oscillatoria</i>	0.09	0.26	0.64
<i>Dolichospermum</i>	0.10	0.20	0.99
<i>Pseudanabaena</i>	0.03	0.20	0.25
<i>Phormidium</i>	0.09	0.20	0.31
<i>Lyngbya</i>	0.05	0.15	0.27

31 The table above exhibits the growth rates of 14 typical cyanobacterial genera based on 1480
 32 records from the references, where p_{10} , p_{50} and p_{90} represent the 10th, 50th and 90th
 33 quantile values of the corresponding cyanobacterial genera growth rates, respectively. The raw
 34 data for these cyanobacterial growth rates and the corresponding references are summarised
 35 in an additional excel sheet.

Table 7 Main water quality parameters in ZXD reservoir during the investigation period (April 2021, n = 71), all values are expressed as mean values with standard deviations, variance analysis were performed to compare the differences.

Parameter	Bottom	Middle	Surface	p-value
MIB _t (ng L ⁻¹)	13.2 ± 7.6	15.5 ± 10.3	17.2 ± 10.3	p = 0.3651
GSM _t (ng L ⁻¹)	4.6 ± 0.9	4.7 ± 0.2	4.8 ± 0.2	p = 0.3885
MIB _d (ng L ⁻¹)	10.0 ± 5.8	11.3 ± 5.4	11.7 ± 5.7	p = 0.5733
GSM _d (ng L ⁻¹)	6.3 ± 1.0	6.2 ± 0.8	6.2 ± 0.9	p = 0.9785
TN (μg L ⁻¹)	1587 ± 165	1629 ± 133	1626 ± 132	p = 0.5374
TP (μg L ⁻¹)	15 ± 4	15 ± 3	15 ± 2	p = 0.7628
⁶¹ NH ₄ -N (μg L ⁻¹)	80 ± 71	67 ± 56	73 ± 50	p = 0.7645
NO ₃ -N (μg L ⁻¹)	1438 ± 153	1441 ± 133	1435 ± 142	p = 0.9866
pH	8.4 ± 0.1	8.4 ± 0.1	8.4 ± 0.1	p = 0.9002
DO (mg L ⁻¹)	5.8 ± 2.2	7.7 ± 0.8	8.0 ± 0.5	p < 0.0001
Turb. (NTU)	17.2 ± 39.8	5.5 ± 6.3	4.7 ± 5.4	p = 0.1245
Temp. (°C)	21.5 ± 1.0	22.2 ± 0.8	22.8 ± 0.8	p < 0.0001
Salinity (psu)	0.16 ± 0.01	0.16 ± 0.01	0.16 ± 0.01	p = 0.5897
Cond. (μs cm ⁻¹)	342.9 ± 20.4	337.4 ± 15.4	337.9 ± 13.9	p = 0.4739
TDS (mg L ⁻¹)	221.5 ± 13.1	218.1 ± 10.0	218.8 ± 8.9	p = 0.5300
Chl <i>a</i> (μg L ⁻¹)	7.3 ± 14.8	7.8 ± 13.1	7.3 ± 6.3	p = 0.9861

Parameter	Bottom	Middle	Surface	<i>p</i>-value
PC ($\mu\text{g L}^{-1}$)	0.1 ± 0.22	0.2 ± 0.2	0.3 ± 0.2	$p = 0.0191$
ORP (mv)	-75.4 ± 214.4	-75.9 ± 214.4	-81.3 ± 216.0	$p = 0.9946$

Table 8 Main water quality in surface layer ZXD reservoir during the investigation period (April 2021, n = 71), all values are expressed as mean values with standard deviations, variance analysis was performed to compare the differences.

Parameter	ZXD01	ZXD02	ZXD03	ZXD04	ZXD05	ZXD06	<i>p</i> -value
MIB _t (ng L ⁻¹)	16.3 ± 9.0	12.1 ± 9.2	16.9 ± 8.8	11.8 ± 11.0	16.4 ± 9.3	18.2 ± 9.8	<i>p</i> = 0.4704
GSM _t (ng L ⁻¹)	4.8 ± 0.3	4.7 ± 0.1	4.7 ± 0.1	4.4 ± 1.1	4.8 ± 0.2	4.7 ± 0.2	<i>p</i> = 0.3951
MIB _d (ng L ⁻¹)	11.1 ± 4.0	8.9 ± 5.9	11.2 ± 5.3	10.5 ± 7.3	12.4 ± 6.3	12.0 ± 4.8	<i>p</i> = 0.7544
GSM _d (ng L ⁻¹)	6.9 ± 0.5	5.8 ± 0.7	5.9 ± 0.8	6.6 ± 1.0	6.2 ± 0.8	6.0 ± 1.1	<i>p</i> = 0.0105
TN (µg L ⁻¹)	1620 ± 62	1649 ± 136	1548 ± 137	1640 ± 203	1608 ± 160	1625 ± 131	<i>p</i> = 0.6064
TP (µg L ⁻¹)	15 ± 3	15 ± 3	14 ± 4	14 ± 3	16 ± 3	15 ± 3	<i>p</i> = 0.8021
²¹ NH ₄ -N (µg L ⁻¹)	67 ± 59	71 ± 76	70 ± 59	81 ± 45	75 ± 67	74 ± 57	<i>p</i> = 0.9955
NO ₃ -N (µg L ⁻¹)	1422 ± 63	1491 ± 110	1366 ± 166	1495 ± 189	1442 ± 145	1418 ± 120	<i>p</i> = 0.2115
pH	8.4 ± 0.1	8.4 ± 0.1	8.4 ± 0.1	8.4 ± 0.1	8.4 ± 0.1	8.4 ± 0.1	<i>p</i> = 0.9492
DO (mg L ⁻¹)	7.6 ± 0.8	7.2 ± 1.3	6.1 ± 2.8	7.3 ± 1.3	7.7 ± 0.9	7.2 ± 1.7	<i>p</i> = 0.1643
Turb. (NTU)	3.2 ± 2.0	29.5 ± 55.1	3.0 ± 2.6	7.9 ± 9.9	4.9 ± 5.4	7.4 ± 10.0	<i>p</i> = 0.0593
Temp. (°C)	22.3 ± 0.8	22.2 ± 1.4	21.8 ± 1.2	21.9 ± 0.9	22.5 ± 0.8	22.2 ± 0.9	<i>p</i> = 0.6057
Salinity (psu)	0.16 ± 0.01	0.16 ± 0.01	0.17 ± 0.01	0.16 ± 0.01	0.16 ± 0.01	0.16 ± 0.01	<i>p</i> = 0.4115
Cond. (µs cm ⁻¹)	332.0 ± 18.2	343.1 ± 17.2	344.6 ± 19.1	336.3 ± 16.2	341.4 ± 14.0	339.2 ± 15.2	<i>p</i> = 0.4689
TDS (mg L ⁻¹)	214.8 ± 11.6	221.7 ± 10.8	222.7 ± 12.1	217.3 ± 10.8	220.8 ± 8.9	219.3 ± 9.9	<i>p</i> = 0.4936
Chl <i>a</i> (µg L ⁻¹)	5.5 ± 3.9	18.4 ± 25.8	5.6 ± 5.5	4.2 ± 4.4	5.5 ± 4.8	6.6 ± 6.4	<i>p</i> = 0.0352

Parameter	ZXD01	ZXD02	ZXD03	ZXD04	ZXD05	ZXD06	<i>p</i> -value
PC ($\mu\text{g L}^{-1}$)	0.16 \pm 0.16	0.30 \pm 0.35	0.21 \pm 0.21	0.17 \pm 0.18	0.21 \pm 0.17	0.22 \pm 0.19	<i>p</i> = 0.6865
ORP (mv)	-111.9 \pm 232.9	-88.6 \pm 228.1	-78.2 \pm 223.5	-62.3 \pm 209.4	-69.5 \pm 213.1	-55.7 \pm 206.5	<i>p</i> = 0.9911
SD (cm)	163.3 \pm 70.9	151.5 \pm 69.8	160.0 \pm 82.8	168.8 \pm 106.9	150.0 \pm 86.8	166.3 \pm 76.0	<i>p</i> = 0.9993
z_{max} (m)	6.4 \pm 1.3	4.5 \pm 1.4	11.0 \pm 0.5	6.7 \pm 0.5	4.9 \pm 0.3	7.0 \pm 0.8	<i>p</i> < 0.0001

Table 9 Main water quality variation before and after construction in reservoir (considering all three layers, n = 71), all values are expressed as mean values with standard deviations, variance analysis were performed to compare the differences.

Parameter	Before	After	p-value
MIB _t (ng L ⁻¹)	22.2 ± 8.7	8.6 ± 3.7	p < 0.0001
GSM _t (ng L ⁻¹)	4.6 ± 0.8	4.7 ± 0.1	p = 0.4035
MIB _d (ng L ⁻¹)	14.8 ± 5.4	7.4 ± 2.5	p < 0.0001
GSM _d (ng L ⁻¹)	6.3 ± 1.0	6.2 ± 0.82	p = 0.9307
TN (µg L ⁻¹)	1563 ± 97	1664 ± 163	p = 0.0023
TP (µg L ⁻¹)	14 ± 2	15 ± 3	p = 0.1520
23 NH ₄ -N (µg L ⁻¹)	98 ± 60	48 ± 46	p = 0.0002
NO ₃ -N (µg L ⁻¹)	1368 ± 118	1507 ± 128	p < 0.0001
pH	8.4 ± 0.1	8.3 ± 0.1	p < 0.0001
DO (mg L ⁻¹)	6.8 ± 2.2	7.5 ± 0.9	p = 0.0661
Turb. (NTU)	6.0 ± 11.4	12.0 ± 31.0	p = 0.2885
Temp. (°C)	21.9 ± 1.1	22.4 ± 0.9	p = 0.0490
Salinity (psu)	0.17 ± 0.01	0.16 ± 0.01	p < 0.0001
Cond. (µs cm ⁻¹)	351.5 ± 12.4	327.6 ± 10.9	p < 0.0001
TDS (mg L ⁻¹)	227.2 ± 7.8	211.9 ± 7.1	p < 0.0001
Chl a (µg L ⁻¹)	11.3 ± 14.5	3.7 ± 6.5	p = 0.0053

Parameter	Before	After	<i>p</i>-value
PC ($\mu\text{g L}^{-1}$)	0.30 \pm 0.25	0.12 \pm 0.11	<i>p</i> = 0.0001
ORP (mv)	41.3 \pm 18.6	-193.2 \pm 248	<i>p</i> < 0.0001
SD (cm)	157.5 \pm 19.1	162.4 \pm 105.1	<i>p</i> = 0.8748
z_{mix} (m)	4.0 \pm 0.9	4.7 \pm 2.8	<i>p</i> = 0.7552
z_{max} (m)	6.7 \pm 2.6	6.8 \pm 2.1	<i>p</i> = 0.9401
HRT (day)	18.0 \pm 2.2	5.4 \pm 0.8	<i>p</i> < 0.0001

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Aphanizomenon aphanizomenoides</i>			28.8	0.36	(Mehnert et al., 2010)
<i>Aphanizomenon flos-aquae</i>			25.0	1.11	(Foy et al., 1976)
<i>Aphanizomenon flos-aquae</i>			22.0	0.16	(Lehtimaki et al., 1997)
<i>Aphanizomenon flos-aquae</i>			24.7	0.27	(Mehnert et al., 2010)
<i>Aphanizomenon flos-aquae</i>			30.0	0.22	(Rapala et al., 1993)
<i>Aphanizomenon flos-aquae</i>			23.0	0.90	(Tsujimura et al., 2001)
<i>Aphanizomenon gracile</i>			27.1	0.29	(Mehnert et al., 2010)
<i>Aphanizomenon gracile</i>			20.0	0.42	(Lürling et al., 2013)
<i>Aphanizomenon gracile</i>			25.0	0.58	(Lürling et al., 2013)
<i>Aphanizomenon gracile</i>			37.5	0.81	(Lürling et al., 2013)
<i>Aphanizomenon gracile</i>			12.5	0.75	(Lürling et al., 2013)
<i>Aphanizomenon gracile</i>			27.5	0.87	(Lürling et al., 2013)
<i>Aphanizomenon gracile</i>			37.5	0.85	(Lürling et al., 2013)
<i>Aphanizomenon ovalisporum</i>			32.8	0.36	(Mehnert et al., 2010)
<i>Aphanizomenon</i> sp.			25.0	0.28	(Konopka and Brock, 1978)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	10.0	25.0	0.28	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	22.0	25.0	0.45	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	26.0	25.0	0.54	(Briand et al., 2004)
²⁶ <i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	42.0	25.0	0.57	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	55.0	25.0	0.60	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	60.0	25.0	0.60	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	70.0	25.0	0.65	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	90.0	25.0	0.66	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	110.0	25.0	0.69	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	145.0	25.0	0.62	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	175.0	25.0	0.65	(Briand et al., 2004)
27 <i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	220.0	25.0	0.63	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	295.0	25.0	0.61	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ACT-9502	Z8	390.0	25.0	0.54	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	10.0	20.0	0.20	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	10.0	20.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	10.0	20.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	10.0	28.0	0.22	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	30.0	20.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	30.0	20.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	30.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	30.0	28.0	0.43	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	30.0	28.0	0.42	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	30.0	28.0	0.45	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	50.0	20.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	50.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	50.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	50.0	28.0	0.49	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	50.0	28.0	0.53	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	50.0	28.0	0.47	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	100.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	100.0	20.0	0.30	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	100.0	20.0	0.29	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	100.0	28.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	100.0	28.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C1	Modified JM	100.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	10.0	20.0	0.29	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	10.0	20.0	0.29	(Xiao et al., 2017)
²⁹ <i>Cylindrospermopsis raciborskii</i> C3	Modified JM	10.0	20.0	0.23	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	10.0	28.0	0.22	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	30.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	30.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	30.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	30.0	28.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	30.0	28.0	0.31	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	30.0	28.0	0.26	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	50.0	20.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	50.0	20.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	50.0	20.0	0.44	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	50.0	28.0	0.60	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	50.0	28.0	0.62	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	50.0	28.0	0.69	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	100.0	20.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	100.0	20.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	100.0	20.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	100.0	28.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	100.0	28.0	0.43	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C3	Modified JM	100.0	28.0	0.39	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	10.0	20.0	0.18	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	10.0	20.0	0.15	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	10.0	20.0	0.18	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	10.0	28.0	0.20	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	10.0	28.0	0.20	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	30.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	30.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	30.0	20.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	30.0	28.0	0.31	(Xiao et al., 2017)
³¹ <i>Cylindrospermopsis raciborskii</i> C6	Modified JM	30.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	30.0	28.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	50.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	50.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	50.0	20.0	0.31	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	50.0	28.0	0.38	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	50.0	28.0	0.35	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	50.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	100.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	100.0	20.0	0.32	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	100.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	100.0	28.0	0.44	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	100.0	28.0	0.44	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C6	Modified JM	100.0	28.0	0.47	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	10.0	20.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	10.0	20.0	0.20	(Xiao et al., 2017)
³² <i>Cylindrospermopsis raciborskii</i> C9	Modified JM	10.0	20.0	0.29	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	10.0	28.0	0.20	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	10.0	28.0	0.24	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	10.0	28.0	0.23	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	30.0	28.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	30.0	28.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	30.0	28.0	0.43	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	50.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	50.0	20.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	50.0	20.0	0.36	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	50.0	28.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	50.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> C9	Modified JM	50.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> CIRF-01			20.0	0.40	(Lürling et al., 2013)
<i>Cylindrospermopsis raciborskii</i> CIRF-01			25.0	0.74	(Lürling et al., 2013)
<i>Cylindrospermopsis raciborskii</i> CIRF-01			22.5	0.81	(Lürling et al., 2013)
<i>Cylindrospermopsis raciborskii</i> CIRF-01			27.5	NA	(Lürling et al., 2013)
<i>Cylindrospermopsis raciborskii</i> CIRF-01			40.0	0.79	(Lürling et al., 2013)
<i>Cylindrospermopsis raciborskii</i> CIRF-01			15.0	0.72	(Lürling et al., 2013)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	20.0	0.56	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	25.0	0.98	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	30.0	1.04	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	35.0	0.87	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	20.0	0.63	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	25.0	1.20	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	30.0	1.28	(Saker and Griffiths, 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	35.0	0.92	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	20.0	0.49	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	25.0	0.76	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	30.0	0.80	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR1	ASM-1	50.0	35.0	0.81	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	20.0	0.41	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	25.0	1.09	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	30.0	1.00	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	35.0	0.81	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	20.0	0.47	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	25.0	1.30	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	30.0	1.07	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	35.0	0.87	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	20.0	0.33	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	25.0	0.87	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	30.0	0.95	(Saker and Griffiths, 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CR2	ASM-1	50.0	35.0	0.75	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	20.0	0.00	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	25.0	0.45	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	30.0	0.54	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	35.0	0.45	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	20.0	0.00	(Saker and Griffiths, 2000)
³⁵ <i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	25.0	0.70	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	30.0	0.59	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	35.0	0.46	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	20.0	0.00	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	25.0	0.20	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	30.0	0.54	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR3	ASM-1	50.0	35.0	0.46	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	20.0	0.51	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	25.0	0.87	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	30.0	1.06	(Saker and Griffiths, 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	35.0	0.71	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	20.0	0.79	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	25.0	1.23	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	30.0	1.17	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	35.0	0.93	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	20.0	0.24	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	25.0	0.52	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	30.0	0.94	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR4	ASM-1	50.0	35.0	0.48	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	20.0	0.30	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	25.0	0.91	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	30.0	1.22	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	35.0	1.11	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	20.0	0.36	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	25.0	1.07	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	30.0	1.28	(Saker and Griffiths, 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	35.0	1.40	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	20.0	0.24	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	25.0	0.78	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	30.0	1.16	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR5	ASM-1	50.0	35.0	0.81	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	20.0	0.37	(Saker and Griffiths, 2000)
37 <i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	25.0	0.86	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	30.0	1.13	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	35.0	0.90	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	20.0	0.38	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	25.0	1.15	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	30.0	1.32	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	35.0	1.03	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	20.0	0.37	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	25.0	0.57	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	30.0	0.97	(Saker and Griffiths, 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CR6	ASM-1	50.0	35.0	0.75	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	20.0	0.45	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	25.0	0.70	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	30.0	1.18	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	35.0	1.18	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	20.0	0.50	(Saker and Griffiths, 2000)
³⁸ <i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	25.0	0.70	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	30.0	1.39	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	35.0	1.49	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	20.0	0.39	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	25.0	0.69	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	30.0	0.96	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CR7	ASM-1	50.0	35.0	0.77	(Saker and Griffiths, 2000)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	11.0	25.0	0.33	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	22.0	25.0	0.49	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	38.0	25.0	0.61	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	45.0	25.0	0.62	(Briand et al., 2004)
³⁹ <i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	52.0	25.0	0.64	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	58.0	25.0	0.73	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	60.0	25.0	0.60	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	68.0	25.0	0.69	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	135.0	25.0	0.73	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	160.0	25.0	0.63	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	211.0	25.0	0.59	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	265.0	25.0	0.70	(Briand et al., 2004)
40 <i>Cylindrospermopsis raciborskii</i> CYP-026J	Z8	380.0	25.0	0.60	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	10.0	25.0	0.33	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	11.0	25.0	0.26	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	22.0	25.0	0.47	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	50.0	25.0	0.73	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	70.0	25.0	0.73	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	90.0	25.0	0.74	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	110.0	25.0	0.79	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	145.0	25.0	0.71	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	180.0	25.0	0.74	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	225.0	25.0	0.70	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	290.0	25.0	0.70	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> ITEP-A3	Z8	400.0	25.0	0.56	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	5.0	26.0	0.12	(Bonilla et al., 2012)
⁴¹ <i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	15.0	26.0	0.35	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	35.0	26.0	0.49	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	45.0	26.0	0.53	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	60.0	26.0	0.54	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	80.0	26.0	0.59	(Bonilla et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	78.0	26.0	0.65	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	100.0	26.0	0.71	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	125.0	26.0	0.39	(Bonilla et al., 2012)
⁴² <i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	150.0	26.0	0.73	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	5.0	26.0	0.23	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	15.0	26.0	0.45	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	35.0	26.0	0.53	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	70.0	26.0	0.50	(Bonilla et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	80.0	26.0	0.69	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	78.0	26.0	0.87	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	100.0	26.0	0.59	(Bonilla et al., 2012)
⁴³ <i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	110.0	26.0	0.51	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	150.0	26.0	0.46	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	5.0	26.0	0.14	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	15.0	26.0	0.40	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	35.0	26.0	0.58	(Bonilla et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	45.0	26.0	0.45	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	75.0	26.0	0.65	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	80.0	26.0	0.76	(Bonilla et al., 2012)
⁴⁴ <i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	125.0	26.0	0.38	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	100.0	26.0	0.74	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	70.0	26.0	0.55	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	145.0	26.0	0.53	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	5.0	26.0	0.18	(Bonilla et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	35.0	26.0	0.52	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	45.0	26.0	0.46	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	90.0	26.0	0.57	(Bonilla et al., 2012)
⁴⁵ <i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	100.0	26.0	0.72	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	120.0	26.0	0.73	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> MVCC14	BG11	145.0	26.0	0.50	(Bonilla et al., 2012)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	10.0	25.0	0.26	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	15.0	25.0	0.33	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	22.0	25.0	0.47	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	32.0	25.0	0.39	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	36.0	25.0	0.59	(Briand et al., 2004)
⁴⁶ <i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	40.0	25.0	0.45	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	85.0	25.0	0.45	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	117.0	25.0	0.50	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	145.0	25.0	0.54	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	178.0	25.0	0.56	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	210.0	25.0	0.57	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	245.0	25.0	0.56	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	290.0	25.0	0.57	(Briand et al., 2004)
⁴⁷ <i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	360.0	25.0	0.55	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	400.0	25.0	0.56	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC117.02	Z8	486.0	25.0	0.54	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	10.0	25.0	0.37	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	22.0	25.0	0.50	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	26.0	25.0	0.64	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	42.0	25.0	0.67	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	51.0	25.0	0.68	(Briand et al., 2004)
⁴⁸ <i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	55.0	25.0	0.68	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	75.0	25.0	0.70	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	86.0	25.0	0.72	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	110.0	25.0	0.64	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	125.0	25.0	0.64	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	165.0	25.0	0.68	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	200.0	25.0	0.65	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	280.0	25.0	0.64	(Briand et al., 2004)
⁴⁹ <i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	380.0	25.0	0.54	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	30.0	15.0	0.08	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	30.0	20.0	0.25	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	30.0	25.0	0.41	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	30.0	30.0	0.42	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	30.0	35.0	0.38	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC98.14	Z8	30.0	40.0	0.17	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	10.0	25.0	0.32	(Briand et al., 2004)
50 <i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	22.0	25.0	0.44	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	42.0	25.0	0.53	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	26.0	25.0	0.49	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	51.0	25.0	0.54	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	60.0	25.0	0.56	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	90.0	25.0	0.56	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	110.0	25.0	0.58	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	125.0	25.0	0.64	(Briand et al., 2004)
⁵¹ <i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	140.0	25.0	0.56	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	175.0	25.0	0.63	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	210.0	25.0	0.62	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	275.0	25.0	0.59	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i> PMC99.12	Z8	365.0	25.0	0.57	(Briand et al., 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WC01	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC01	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC01	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC02	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC02	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC02	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)
52 <i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	15.0	28.0	0.22	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	15.0	28.0	0.21	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	15.0	28.0	0.20	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	10.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	10.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	10.0	20.0	0.29	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	30.0	20.0	0.18	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	30.0	20.0	0.20	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	30.0	20.0	0.19	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	10.0	28.0	0.23	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	10.0	28.0	0.23	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	10.0	28.0	0.24	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	30.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	30.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	30.0	28.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	50.0	20.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	50.0	20.0	0.38	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	50.0	20.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	50.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	50.0	28.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	50.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	100.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	100.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	100.0	20.0	0.22	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	100.0	28.0	0.43	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	100.0	28.0	0.44	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WC03	Modified JM	100.0	28.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC04	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC04	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC04	Modified JM	15.0	28.0	0.19	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
54 <i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	10.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	10.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	10.0	20.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	30.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	30.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	30.0	20.0	0.24	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	10.0	28.0	0.22	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	10.0	28.0	0.22	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	10.0	28.0	0.22	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	30.0	28.0	0.29	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	30.0	28.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	30.0	28.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	50.0	20.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	50.0	20.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	50.0	20.0	0.24	(Xiao et al., 2017)
⁵⁵ <i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	50.0	28.0	0.25	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	50.0	28.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	50.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	100.0	20.0	0.39	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	100.0	20.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	100.0	20.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	100.0	28.0	0.39	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	100.0	28.0	0.38	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC05	Modified JM	100.0	28.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC06	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WC06	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC06	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC07	Modified JM	15.0	28.0	0.22	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC07	Modified JM	15.0	28.0	0.21	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC07	Modified JM	15.0	28.0	0.20	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	10.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	10.0	20.0	0.31	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	10.0	20.0	0.31	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	30.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	30.0	20.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	30.0	20.0	0.35	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	10.0	28.0	0.25	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	10.0	28.0	0.22	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	10.0	28.0	0.20	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	30.0	28.0	0.35	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	30.0	28.0	0.36	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	30.0	28.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	50.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	50.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	50.0	20.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	50.0	28.0	0.35	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	50.0	28.0	0.29	(Xiao et al., 2017)
57 <i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	50.0	28.0	0.38	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	100.0	20.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	100.0	20.0	0.20	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	100.0	20.0	0.22	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	100.0	28.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	100.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WC09	Modified JM	100.0	28.0	0.35	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	15.0	28.0	0.12	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	15.0	28.0	0.13	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	15.0	28.0	0.11	(Willis et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	10.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	10.0	20.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	10.0	20.0	0.19	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	10.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	10.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	10.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	30.0	20.0	0.21	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	30.0	20.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	30.0	20.0	0.28	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	30.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	30.0	28.0	0.40	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	30.0	28.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	50.0	20.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	50.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	50.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	50.0	28.0	0.36	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	50.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	50.0	28.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	100.0	20.0	0.45	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	100.0	20.0	0.46	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	100.0	20.0	0.45	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	100.0	28.0	0.39	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	100.0	28.0	0.44	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS01	Modified JM	100.0	28.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS02	Modified JM	15.0	28.0	0.10	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS02	Modified JM	15.0	28.0	0.12	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS02	Modified JM	15.0	28.0	0.08	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS03	Modified JM	15.0	28.0	0.13	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS03	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS03	Modified JM	15.0	28.0	0.21	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS04	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS04	Modified JM	15.0	28.0	0.21	(Willis et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WS04	Modified JM	15.0	28.0	0.09	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	10.0	20.0	0.17	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	10.0	20.0	0.16	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	10.0	20.0	0.17	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	10.0	28.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	10.0	28.0	0.34	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	10.0	28.0	0.24	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	30.0	20.0	0.27	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	30.0	20.0	0.26	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	30.0	20.0	0.29	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	30.0	28.0	0.33	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	30.0	28.0	0.37	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	30.0	28.0	0.42	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	50.0	20.0	0.31	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	50.0	20.0	0.30	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	50.0	20.0	0.32	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	50.0	28.0	0.42	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	50.0	28.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	50.0	28.0	0.41	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	100.0	20.0	0.35	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	100.0	20.0	0.38	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	100.0	20.0	0.36	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	100.0	28.0	0.49	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	100.0	28.0	0.51	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS05	Modified JM	100.0	28.0	0.51	(Xiao et al., 2017)
<i>Cylindrospermopsis raciborskii</i> WS06	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS06	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS06	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS07	Modified JM	15.0	28.0	0.10	(Willis et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WS07	Modified JM	15.0	28.0	0.11	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS07	Modified JM	15.0	28.0	0.12	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS08	Modified JM	15.0	28.0	0.11	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS08	Modified JM	15.0	28.0	0.12	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS08	Modified JM	15.0	28.0	0.10	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS09	Modified JM	15.0	28.0	0.12	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS09	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS09	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS10	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS10	Modified JM	15.0	28.0	0.13	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS10	Modified JM	15.0	28.0	0.12	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS11	Modified JM	15.0	28.0	0.13	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS11	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS11	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS12	Modified JM	15.0	28.0	0.19	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS12	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> WS12	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS13	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS13	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS13	Modified JM	15.0	28.0	0.19	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS14	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS14	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS14	Modified JM	15.0	28.0	0.13	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS15	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS15	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS15	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS16	Modified JM	15.0	28.0	0.18	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS16	Modified JM	15.0	28.0	0.17	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS16	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS17	Modified JM	15.0	28.0	0.14	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS17	Modified JM	15.0	28.0	0.15	(Willis et al., 2016)
<i>Cylindrospermopsis raciborskii</i> WS17	Modified JM	15.0	28.0	0.16	(Willis et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	15.0	-0.08	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	15.0	-0.08	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	15.0	-0.10	(Thomas and Litchman, 2015)
64 <i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	15.0	-0.09	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	20.0	0.17	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	20.0	0.17	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	20.0	0.16	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	20.0	0.17	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	25.0	0.40	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	25.0	0.39	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	25.0	0.39	(Thomas and Litchman, 2015)
⁵⁹ <i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	25.0	0.39	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	30.0	0.53	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	30.0	0.52	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	30.0	0.51	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i> <i>floridaD</i>	WC	100.0	30.0	0.52	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii floridaD</i>	WC	100.0	35.0	0.45	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaD</i>	WC	100.0	35.0	0.44	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaD</i>	WC	100.0	35.0	0.43	(Thomas and Litchman, 2015)
⁹⁹ <i>Cylindrospermopsis raciborskii floridaD</i>	WC	100.0	35.0	0.45	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	15.0	-0.04	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	15.0	-0.03	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	15.0	-0.01	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	15.0	-0.02	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	20.0	0.14	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	20.0	0.13	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	20.0	0.12	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	20.0	0.12	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	30.0	0.49	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	30.0	0.47	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	30.0	0.45	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	30.0	0.46	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	40.0	-0.03	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	40.0	-0.05	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	40.0	-0.07	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	40.0	-0.05	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	35.0	0.33	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	35.0	0.32	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	35.0	0.32	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii floridaE</i>	WC	100.0	35.0	0.36	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	15.0	-0.04	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	15.0	-0.02	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	15.0	-0.03	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	15.0	-0.03	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	20.0	0.24	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	20.0	0.23	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	20.0	0.21	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	20.0	0.20	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	25.0	0.38	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	25.0	0.36	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	25.0	0.37	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	25.0	0.36	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	35.0	0.53	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	35.0	0.50	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	35.0	0.45	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	35.0	0.48	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	30.0	0.51	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	30.0	0.50	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	30.0	0.48	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	30.0	0.49	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	40.0	-0.10	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	40.0	-0.16	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	40.0	-0.13	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii indiana</i>	WC	100.0	40.0	-0.12	(Thomas and Litchman, 2015)
<i>Cylindrospermopsis raciborskii</i>			30.0	0.00	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			29.0	0.60	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			31.0	0.70	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			30.0	0.40	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			29.0	0.60	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			29.0	0.80	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			29.0	0.70	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			31.0	0.40	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			30.0	0.50	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			30.0	0.50	(Briand et al., 2004)
<i>Cylindrospermopsis raciborskii</i>			27.8	0.34	(Mehnert et al., 2010)
<i>Cylindrospermopsis raciborskii</i>			28.0	0.58	(Saker et al., 1999)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Cylindrospermopsis raciborskii</i>			32.0	0.71	(Soares et al., 2010)
<i>Cylindrospermopsis raciborskii</i>		133.4 \pm 13.5	31.3 \pm 19.0	0.77	(Xiao et al., 2020)
<i>Dolichospermum aphanizomenoides</i>			35.0	1.46	(Sabour et al., 2009)
<i>Dolichospermum bergii</i>			26.5	0.25	(Mehnert et al., 2010)
<i>Dolichospermum circinalis Ana318</i>	ASM	100.0	25.0	0.12	(Li et al., 2012)
<i>Dolichospermum circinalis Ana318</i>	ASM	10.0	25.0	0.11	(Li et al., 2012)
<i>Dolichospermum circinalis Ana318</i>	ASM	100.0	18.0	0.13	(Li et al., 2012)
<i>Dolichospermum circinalis Ana318</i>	ASM	10.0	18.0	0.12	(Li et al., 2012)
<i>Dolichospermum flos-aquae</i>			20.0	0.78	(Foy et al., 1976)
<i>Dolichospermum flos-aquae</i>			39.0	1.01	(Novak and Brune, 1985)
<i>Dolichospermum flos-aquae</i>			20.0	0.20	(Rapala et al., 1993)
<i>Dolichospermum flos-aquae</i>			20.0	0.19	(Rapala et al., 1993)
<i>Dolichospermum macrospora</i>			25.5	0.19	(Mehnert et al., 2010)
<i>Dolichospermum mendotae</i>			20.0	0.20	(Rapala et al., 1993)
<i>Dolichospermum smithii</i>	CT	60.0	28.0	0.21	(Shen et al., 2020)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Dolichospermum smithii</i>	CT(N elevated)	60.0	28.0	0.22	(Shen et al., 2020)
<i>Dolichospermum spiroides</i>			24.0	0.77	(Seki et al., 1981)
<i>Dolichospermum ucrainica</i> CHAB 2155	CT	30.0	10.0	-0.04	(Wang and Li, 2015)
<i>Dolichospermum ucrainica</i> CHAB 2155	CT	30.0	25.0	0.09	(Wang and Li, 2015)
<i>Dolichospermum ucrainica</i> CHAB 2155	CT	30.0	35.0	0.10	(Wang and Li, 2015)
<i>Dolichospermum ucrainica</i> CHAB 2155	CT	10.0	25.0	0.08	(Wang and Li, 2015)
¹⁷ <i>Dolichospermum ucrainica</i> CHAB 2155	CT	60.0	25.0	0.10	(Wang and Li, 2015)
<i>Dolichospermum ukrainica</i>			26.0	0.78	(Tsujiura and Okubo, 2003)
<i>Dolichospermum variabilis</i>			35.0	1.20	(Kratz and Myers, 1955)
<i>Dolichospermum variabilis</i>			30.0	0.15	(Wang et al., 2007)
<i>Dolichospermum</i> sp.			25.0	0.13	(Konopka and Brock, 1978)
<i>Dolichospermum</i> sp.			32.0	1.25	(Nalewajko and Murphy, 2001)
<i>Dolichospermum</i> sp.			20.0	0.15	(Rapala et al., 1997)
<i>Dolichospermum</i> sp.			25.0	0.14	(Rapala et al., 1997)
<i>Dolichospermum</i> sp.			28.0	0.80	(Vincent and Silvester, 1979)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Dolichospermum sp. PCC7122</i>			20.0	0.46	(Lürling et al., 2013)
<i>Dolichospermum sp. PCC7122</i>			25.0	0.93	(Lürling et al., 2013)
<i>Dolichospermum sp. PCC7122</i>			27.5	0.75	(Lürling et al., 2013)
<i>Dolichospermum sp. PCC7122</i>			30.0	NA	(Lürling et al., 2013)
<i>Dolichospermum sp. PCC7122</i>			32.5	0.53	(Lürling et al., 2013)
<i>Dolichospermum sp. PCC7122</i>			35.0	0.58	(Lürling et al., 2013)
⁷² <i>Limnothrix redekei</i>	BG11	33.8 ~ 54.1	28.0	0.15	(Tiwari et al., 2001)
<i>Limnothrix redekei</i>	MV-NH4	120.0	10.0	0.17	(Nicklisch, 1999)
<i>Limnothrix redekei</i>	MV-NH4	120.0	15.0	0.53	(Nicklisch, 1999)
<i>Limnothrix redekei</i>				0.10	(Nicklisch, 1992)
<i>Limnothrix redekei</i>		24.0	15.0	0.52	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>		32.0	15.0	0.37	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>		27.0	15.0	0.31	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>			15.0	0.21	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>			15.0	0.22	(Shatwell et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Limnothrix redekei</i>			10.0	0.32	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>			20.0	0.70	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>			10.0	0.22	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>			20.0	0.51	(Shatwell et al., 2012)
<i>Limnothrix redekei</i>	Modified MIV	122.0	20.0	0.47	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	75.0	20.0	0.37	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	122.0	20.0	0.53	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	134.0	20.0	0.80	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	56.0	20.0	0.70	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	254.0	20.0	0.50	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	60.0	20.0	1.11	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	183.0	20.0	0.69	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	270.0	20.0	0.70	(Nicklisch, 1998)
<i>Limnothrix redekei</i>	Modified MIV	427.0	20.0	0.28	(Nicklisch, 1998)
<i>Lyngbya dendrobia</i>	BG11	33.8 ~ 54.1	28.0	0.06	(Tiwari et al., 2001)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Lyngbya kuetzinghii</i>			25.0	0.14	(Zhang et al., 2009)
<i>Lyngbya lachneri</i>	BG11	33.8 ~ 54.1	28.0	0.08	(Tiwari et al., 2001)
<i>Lyngbya majuscula</i>				0.33	(Elmetri and Bell, 2004)
<i>Lyngbya majuscula</i>				0.25	(Elmetri and Bell, 2004)
<i>Lyngbya majuscula</i>				0.19	(Elmetri and Bell, 2004)
⁷⁴ <i>Lyngbya majuscula</i>				0.14	(Elmetri and Bell, 2004)
<i>Lyngbya majuscula</i>				0.04	(Elmetri and Bell, 2004)
<i>Lyngbya majuscula</i>				0.05	(Elmetri and Bell, 2004)
<i>Lyngbya nigra</i>	BG11	33.8 ~ 54.1	28.0	0.18	(Tiwari et al., 2001)
<i>Lyngbya palmarum</i>	BG11	33.8 ~ 54.1	28.0	0.28	(Tiwari et al., 2001)
<i>Lyngbya spiralis</i>	BG11	33.8 ~ 54.1	28.0	0.22	(Tiwari et al., 2001)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Lyngbya truncicola</i>	BG11	33.8 ~ 54.1	28.0	0.12	(Tiwari et al., 2001)
<i>Microcoleus chthonoplastes MPI CHI-1</i>	BG11	35.0	25.0	0.55	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI CHI-1</i>	BG11	35.0	25.0	0.47	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI CHI-1</i>	BG11	35.0	25.0	0.28	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI</i> EBD-1	BG11	35.0	25.0	0.24	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI</i> EBD-1	BG11	35.0	25.0	0.26	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI</i> EBD-1	BG11	35.0	25.0	0.18	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI</i> GN5-1	BG11	35.0	25.0	0.21	(Karsten, 1996)
<i>Microcoleus chthonoplastes MPI</i> GN5-1	BG11	35.0	25.0	0.23	(Karsten, 1996)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcoleus chthonoplastes</i> MPI GN5-1	BG11	35.0	25.0	0.17	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI GNL-1	BG11	35.0	25.0	0.23	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI GNL-1	BG11	35.0	25.0	0.29	(Karsten, 1996)
76 <i>Microcoleus chthonoplastes</i> MPI GNL-1	BG11	35.0	25.0	0.24	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI SOL-1	BG11	35.0	25.0	0.13	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI SOL-1	BG11	35.0	25.0	0.17	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI SOL-1	BG11	35.0	25.0	0.20	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI SPW-1	BG11	35.0	25.0	0.29	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> MPI SPW-1	BG11	35.0	25.0	0.29	(Karsten, 1996)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcoleus chthonoplastes</i> MPI	BG11	35.0	25.0	0.18	(Karsten, 1996)
SPW-1					
<i>Microcoleus chthonoplastes</i> MPI	BG11	35.0	25.0	0.35	(Karsten, 1996)
TOW-1					
<i>Microcoleus chthonoplastes</i> MPI	BG11	35.0	25.0	0.35	(Karsten, 1996)
TOW-1					
77 <i>Microcoleus chthonoplastes</i> MPI	BG11	35.0	25.0	0.33	(Karsten, 1996)
TOW-1					
<i>Microcoleus chthonoplastes</i> PCC 7420	BG11	35.0	25.0	0.42	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> PCC 7420	BG11	35.0	25.0	0.46	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> PCC 7420	BG11	35.0	25.0	0.21	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> UBM HID	BG11	35.0	25.0	0.42	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> UBM HID	BG11	35.0	25.0	0.35	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> UBM HID	BG11	35.0	25.0	0.25	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> UBM WIS	BG11	35.0	25.0	0.54	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i> UBM WIS	BG11	35.0	25.0	0.46	(Karsten, 1996)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcoleus chthonoplastes</i> UBM WIS	BG11	35.0	25.0	0.16	(Karsten, 1996)
<i>Microcoleus chthonoplastes</i>	BG11	33.8 ~ 54.1	28.0	0.20	(Tiwari et al., 2001)
<i>Microcoleus paludosus</i>	BG11	33.8 ~ 54.1	28.0	0.17	(Tiwari et al., 2001)
<i>Microcoleus</i> spp.				0.03	(Lababpour and Kaviani, 2016)
<i>Microcoleus</i> spp.				0.06	(Lababpour and Kaviani, 2016)
<i>Microcoleus steenstrupii</i> HS024				3.22	(Giraldo-Silva et al., 2019)
<i>Microcoleus steenstrupii</i> HS024				0.30	(Giraldo-Silva et al., 2019)
<i>Microcoleus steenstrupii</i> HS024				0.30	(Giraldo-Silva et al., 2019)
<i>Microcoleus steenstrupii</i> JS010				1.37	(Giraldo-Silva et al., 2019)
<i>Microcoleus steenstrupii</i> JS010				-1.00	(Giraldo-Silva et al., 2019)
<i>Microcoleus steenstrupii</i> JS010				14.80	(Giraldo-Silva et al., 2019)
<i>Microcoleus vaginatus</i> FB020				1.17	(Giraldo-Silva et al., 2019)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcoleus vaginatus</i> FB020				1.80	(Giraldo-Silva et al., 2019)
<i>Microcoleus vaginatus</i> FB020				0.50	(Giraldo-Silva et al., 2019)
<i>Microcoleus vaginatus</i> HSN003				2.12	(Giraldo-Silva et al., 2019)
<i>Microcoleus vaginatus</i> HSN003				0.60	(Giraldo-Silva et al., 2019)
<i>Microcoleus vaginatus</i> HSN003				0.30	(Giraldo-Silva et al., 2019)
<i>Microcoleus vaginatus</i>			21.0	0.22	(Novak and Brune, 1985)
<i>Microcystis aeruginosa</i> CYA140			20.0	0.26	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> CYA140			25.0	0.77	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> CYA140			22.5	0.82	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> CYA140			27.5	0.94	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> CYA140			25.0	0.93	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> CYA140			30.0	0.70	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	15.0	0.29	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	15.0	0.31	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	15.0	0.30	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	20.0	0.51	(Marinho et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	20.0	0.53	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	20.0	0.54	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	25.0	0.56	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	25.0	0.55	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	25.0	0.56	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	30.0	0.72	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	30.0	0.68	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	30.0	0.66	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	35.0	0.79	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	35.0	0.77	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	35.0	0.78	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	10.0	25.0	0.33	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	10.0	25.0	0.31	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	10.0	25.0	0.29	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	15.0	25.0	0.35	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	15.0	25.0	0.33	(Marinho et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB469	BG11	15.0	25.0	0.35	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	25.0	0.43	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	25.0	0.42	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	25.0	0.43	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	25.0	0.62	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	25.0	0.62	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	25.0	0.63	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	25.0	0.63	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	25.0	0.63	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	25.0	0.62	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	25.0	0.63	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	25.0	0.62	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	25.0	0.63	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	25.0	0.69	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	25.0	0.70	(Marinho et al., 2013)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	25.0	0.69	(Marinho et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	20.0	0.33	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	20.0	0.34	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	20.0	0.32	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	20.0	0.41	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	20.0	0.39	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	20.0	0.37	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	35.0	20.0	0.42	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	35.0	20.0	0.41	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	35.0	20.0	0.40	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	20.0	0.44	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	20.0	0.46	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	20.0	0.43	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	45.0	20.0	0.52	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	45.0	20.0	0.54	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	45.0	20.0	0.50	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	20.0	0.53	(Li et al., 2014)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	20.0	0.55	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	20.0	0.51	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	20.0	0.56	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	20.0	0.58	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	20.0	0.53	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	20.0	0.56	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	20.0	0.55	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	20.0	0.58	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	25.0	0.52	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	25.0	0.54	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	25.0	25.0	0.51	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	25.0	0.54	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	25.0	0.56	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	30.0	25.0	0.53	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	35.0	25.0	0.56	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	35.0	25.0	0.57	(Li et al., 2014)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB469	BG11	35.0	25.0	0.54	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	25.0	0.60	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	25.0	0.61	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	40.0	25.0	0.58	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	45.0	25.0	0.60	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	45.0	25.0	0.61	(Li et al., 2014)
84 <i>Microcystis aeruginosa</i> FACHB469	BG11	45.0	25.0	0.59	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	25.0	0.61	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	25.0	0.62	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	50.0	25.0	0.60	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	25.0	0.62	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	25.0	0.63	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	55.0	25.0	0.61	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	25.0	0.63	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	25.0	0.65	(Li et al., 2014)
<i>Microcystis aeruginosa</i> FACHB469	BG11	60.0	25.0	0.62	(Li et al., 2014)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB905	Modified BG11	50.0	25.0	0.11	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB905	Modified BG11	50.0	25.0	0.11	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB905	Modified BG11	50.0	25.0	0.10	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB909	Modified BG11	50.0	25.0	0.18	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB909	Modified BG11	50.0	25.0	0.20	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB909	Modified BG11	50.0	25.0	0.15	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB938	Modified BG11	50.0	25.0	0.15	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB938	Modified BG11	50.0	25.0	0.16	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB938	Modified BG11	50.0	25.0	0.14	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB939	Modified BG11	50.0	25.0	0.13	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB939	Modified BG11	50.0	25.0	0.14	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB939	Modified BG11	50.0	25.0	0.13	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB942	Modified BG11	50.0	25.0	0.13	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB942	Modified BG11	50.0	25.0	0.14	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB942	Modified BG11	50.0	25.0	0.12	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB975	Modified BG11	50.0	25.0	0.17	(Shen and Song, 2007)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> FACHB975	Modified BG11	50.0	25.0	0.19	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> FACHB975	Modified BG11	50.0	25.0	0.21	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	15.0	0.05	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	15.0	0.04	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	15.0	0.03	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	15.0	0.04	(Thomas and Litchman, 2015)
⁹⁸ <i>Microcystis aeruginosa</i> GullB00	WC	100.0	20.0	0.16	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	20.0	0.15	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	20.0	0.14	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	20.0	0.15	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	25.0	0.36	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	25.0	0.35	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	25.0	0.34	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	25.0	0.35	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	30.0	0.37	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	30.0	0.35	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	30.0	0.36	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	30.0	0.36	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	35.0	-0.12	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	35.0	-0.14	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	35.0	-0.14	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullB00	WC	100.0	35.0	-0.13	(Thomas and Litchman, 2015)
87 <i>Microcystis aeruginosa</i> GullK00	WC	100.0	15.0	0.06	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	15.0	0.04	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	15.0	0.03	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	15.0	0.04	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	20.0	0.13	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	20.0	0.11	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	20.0	0.10	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	20.0	0.12	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	25.0	0.39	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	25.0	0.38	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	25.0	0.37	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	25.0	0.38	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	30.0	0.40	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	30.0	0.38	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	30.0	0.37	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	30.0	0.38	(Thomas and Litchman, 2015)
∞ <i>Microcystis aeruginosa</i> GullK00	WC	100.0	35.0	-0.14	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	35.0	-0.15	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	35.0	-0.16	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> GullK00	WC	100.0	35.0	-0.15	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> LMECYA	Z8	4.0	20.0	0.07	(Salvador et al., 2016)
<i>Microcystis aeruginosa</i> LMECYA	Z8	20.0	20.0	0.11	(Salvador et al., 2016)
<i>Microcystis aeruginosa</i> LMECYA	Z8	30.0	20.0	0.12	(Salvador et al., 2016)
<i>Microcystis aeruginosa</i> M2	Modified JM	10.0	20.0	0.22	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	10.0	20.0	0.20	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	10.0	20.0	0.21	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> M2	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	30.0	20.0	0.23	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	30.0	20.0	0.23	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	30.0	20.0	0.26	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	30.0	28.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	30.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	30.0	28.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	50.0	20.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	50.0	20.0	0.33	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	50.0	20.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	50.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	50.0	28.0	0.42	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	50.0	28.0	0.42	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	100.0	20.0	0.35	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> M2	Modified JM	100.0	20.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	100.0	20.0	0.39	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	100.0	28.0	0.45	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	100.0	28.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M2	Modified JM	100.0	28.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	10.0	20.0	0.29	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	10.0	20.0	0.26	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	10.0	20.0	0.23	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	10.0	28.0	0.27	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	10.0	28.0	0.28	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	10.0	28.0	0.26	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	30.0	20.0	0.30	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	30.0	20.0	0.32	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	30.0	20.0	0.27	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	30.0	28.0	0.29	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	30.0	28.0	0.29	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> M3	Modified JM	30.0	28.0	0.28	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	50.0	20.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	50.0	20.0	0.38	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	50.0	20.0	0.32	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	50.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	50.0	28.0	0.38	(Xiao et al., 2017)
¹⁶ <i>Microcystis aeruginosa</i> M3	Modified JM	50.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	100.0	20.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	100.0	20.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	100.0	20.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	100.0	28.0	0.31	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	100.0	28.0	0.33	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M3	Modified JM	100.0	28.0	0.34	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	10.0	20.0	0.21	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	10.0	20.0	0.18	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	10.0	20.0	0.20	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> M4	Modified JM	10.0	28.0	0.27	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	10.0	28.0	0.28	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	10.0	28.0	0.25	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	30.0	20.0	0.30	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	30.0	20.0	0.23	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	30.0	20.0	0.27	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	30.0	28.0	0.28	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	30.0	28.0	0.29	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	30.0	28.0	0.26	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	50.0	20.0	0.34	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	50.0	20.0	0.33	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	50.0	20.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	50.0	28.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	50.0	28.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	50.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	100.0	20.0	0.45	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> M4	Modified JM	100.0	20.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	100.0	20.0	0.45	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	100.0	28.0	0.35	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	100.0	28.0	0.32	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M4	Modified JM	100.0	28.0	0.32	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	10.0	20.0	0.27	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	10.0	20.0	0.28	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	10.0	20.0	0.26	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	10.0	28.0	0.16	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	10.0	28.0	0.21	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	10.0	28.0	0.20	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	30.0	20.0	0.24	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	30.0	20.0	0.24	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	30.0	20.0	0.23	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	30.0	28.0	0.31	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	30.0	28.0	0.30	(Xiao et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> M5	Modified JM	30.0	28.0	0.31	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	50.0	28.0	0.41	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	50.0	28.0	0.38	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	50.0	28.0	0.38	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	50.0	20.0	0.27	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	50.0	20.0	0.29	(Xiao et al., 2017)
94 <i>Microcystis aeruginosa</i> M5	Modified JM	50.0	20.0	0.30	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	100.0	20.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	100.0	20.0	0.32	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	100.0	20.0	0.37	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	100.0	28.0	0.31	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	100.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> M5	Modified JM	100.0	28.0	0.36	(Xiao et al., 2017)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	10.0	24.0	0.28	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	10.0	24.0	0.26	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	10.0	24.0	0.26	(Torres et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	60.0	24.0	0.54	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	60.0	24.0	0.53	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	60.0	24.0	0.55	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	100.0	24.0	0.45	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	100.0	24.0	0.45	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	100.0	24.0	0.45	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	500.0	24.0	0.39	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	500.0	24.0	0.41	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	500.0	24.0	0.37	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	40.0	24.0	0.47	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	40.0	24.0	0.46	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-03	Modified WC	40.0	24.0	0.45	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	10.0	24.0	0.29	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	10.0	24.0	0.28	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	10.0	24.0	0.29	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	60.0	24.0	0.45	(Torres et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	60.0	24.0	0.43	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	60.0	24.0	0.47	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	100.0	24.0	0.56	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	100.0	24.0	0.56	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	100.0	24.0	0.56	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	500.0	24.0	0.58	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	500.0	24.0	0.59	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	500.0	24.0	0.56	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	40.0	24.0	0.54	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	40.0	24.0	0.58	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIC-08	Modified WC	40.0	24.0	0.51	(Torres et al., 2016)
<i>Microcystis aeruginosa</i> MIRF	ASM-1	100.0	25.0	0.61	(e. Mello et al., 2012)
<i>Microcystis aeruginosa</i> MIRF	ASM-1	100.0	25.0	0.63	(e. Mello et al., 2012)
<i>Microcystis aeruginosa</i> MIRF	ASM-1	100.0	25.0	0.58	(e. Mello et al., 2012)
<i>Microcystis aeruginosa</i> MT2PCC7806	Modified BG11	39.0	22.0	0.49	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> MT2PCC7806	Modified BG11	39.0	22.0	0.45	(Briand et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> MT2PCC7806	Modified BG11	39.0	22.0	0.53	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> MT2PCC7806	Modified BG11	5.0	22.0	0.11	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> MT2PCC7806	Modified BG11	5.0	22.0	0.10	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> MT2PCC7806	Modified BG11	5.0	22.0	0.12	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	4.0	20.0	0.05	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	4.0	20.0	0.05	(Hesse et al., 2001)
97 <i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	4.0	20.0	0.04	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	9.0	20.0	0.11	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	9.0	20.0	0.10	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	9.0	20.0	0.10	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	11.0	20.0	0.12	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	11.0	20.0	0.11	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	11.0	20.0	0.11	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	20.0	20.0	0.17	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	20.0	20.0	0.15	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	20.0	20.0	0.17	(Hesse et al., 2001)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	30.0	20.0	0.22	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	30.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	30.0	20.0	0.19	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	65.0	20.0	0.21	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	65.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	65.0	20.0	0.19	(Hesse et al., 2001)
⁸⁶ <i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	90.0	20.0	0.21	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	90.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	90.0	20.0	0.19	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	105.0	20.0	0.23	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	105.0	20.0	0.22	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	105.0	20.0	0.21	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	25.0	20.0	0.18	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	25.0	20.0	0.16	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> MTPCC7806	MIV/2	25.0	20.0	0.15	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> Ma17D	BG11	15.0	20.0	0.20	(Bañares-España et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> Ma17D	BG11	15.0	20.0	0.19	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma17D	BG11	15.0	20.0	0.21	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma17D	BG11	176.0	20.0	0.09	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma17D	BG11	176.0	20.0	0.10	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma17D	BG11	176.0	20.0	0.08	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma2M	BG11	15.0	20.0	0.24	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma2M	BG11	15.0	20.0	0.24	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma2M	BG11	15.0	20.0	0.23	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma2M	BG11	176.0	20.0	0.17	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma2M	BG11	176.0	20.0	0.17	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma2M	BG11	176.0	20.0	0.16	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma5D	BG11	15.0	20.0	0.20	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma5D	BG11	15.0	20.0	0.24	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma5D	BG11	15.0	20.0	0.17	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma5D	BG11	176.0	20.0	0.17	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> Ma5D	BG11	176.0	20.0	0.18	(Bañares-España et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> Ma5D	BG11	176.0	20.0	0.17	(Bañares-España et al., 2012)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	20.0	0.26	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	20.0	0.23	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	20.0	0.29	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	25.0	0.77	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	25.0	0.79	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	25.0	0.75	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	27.5	0.82	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	27.5	0.83	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	27.5	0.85	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	30.0	0.94	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	30.0	0.95	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	30.0	0.97	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	32.5	0.93	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	32.5	0.95	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	32.5	0.91	(Lürling et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	35.0	0.70	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	35.0	0.71	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> NIVA-CYA140	Modified WC	80.0	35.0	0.69	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	39.0	22.0	0.46	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	39.0	22.0	0.43	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	39.0	22.0	0.49	(Briand et al., 2012)
101 <i>Microcystis aeruginosa</i> PCC7806	Modified BG11	5.0	22.0	0.12	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	5.0	22.0	0.11	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	5.0	22.0	0.13	(Briand et al., 2012)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	4.0	20.0	0.06	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	4.0	20.0	0.06	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	4.0	20.0	0.07	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	9.0	20.0	0.10	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	9.0	20.0	0.11	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	9.0	20.0	0.10	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	11.0	20.0	0.14	(Hesse et al., 2001)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	11.0	20.0	0.15	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	11.0	20.0	0.14	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	25.0	20.0	0.19	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	25.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	25.0	20.0	0.18	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	35.0	20.0	0.22	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	35.0	20.0	0.23	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	35.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	70.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	70.0	20.0	0.21	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	70.0	20.0	0.19	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	90.0	20.0	0.21	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	90.0	20.0	0.22	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	90.0	20.0	0.20	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	105.0	20.0	0.22	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	105.0	20.0	0.23	(Hesse et al., 2001)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> PCC7806	MIV/2	105.0	20.0	0.21	(Hesse et al., 2001)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	50.0	25.0	0.15	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	50.0	25.0	0.16	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> PCC7806	Modified BG11	50.0	25.0	0.14	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	9.0	22.0	0.20	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	9.0	22.0	0.18	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	9.0	22.0	0.21	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	13.0	22.0	0.17	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	13.0	22.0	0.17	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	13.0	22.0	0.17	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	20.0	22.0	0.26	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	20.0	22.0	0.23	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	20.0	22.0	0.29	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	30.0	22.0	0.28	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	30.0	22.0	0.27	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	30.0	22.0	0.29	(Wiedner et al., 2003)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	38.0	22.0	0.36	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	38.0	22.0	0.34	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	38.0	22.0	0.39	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	43.0	22.0	0.38	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	43.0	22.0	0.35	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	43.0	22.0	0.40	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	65.0	22.0	0.34	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	65.0	22.0	0.34	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	65.0	22.0	0.34	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	125.0	22.0	0.38	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	125.0	22.0	0.36	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	125.0	22.0	0.41	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	150.0	22.0	0.33	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	150.0	22.0	0.28	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	150.0	22.0	0.38	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	250.0	22.0	0.37	(Wiedner et al., 2003)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	250.0	22.0	0.33	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	250.0	22.0	0.41	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	255.0	22.0	0.35	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	255.0	22.0	0.29	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	255.0	22.0	0.42	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	400.0	22.0	0.32	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	400.0	22.0	0.32	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7806	O2-medium	400.0	22.0	0.32	(Wiedner et al., 2003)
<i>Microcystis aeruginosa</i> PCC7820	Modified BG11	50.0	25.0	0.10	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> PCC7820	Modified BG11	50.0	25.0	0.12	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> PCC7820	Modified BG11	50.0	25.0	0.09	(Shen and Song, 2007)
<i>Microcystis aeruginosa</i> PCC7941			20.0	0.58	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941			25.0	0.67	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941			22.5	1.05	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941			27.5	NA	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941			25.0	1.16	(Lürling et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> PCC7941			30.0	1.01	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	20.0	0.58	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	20.0	0.57	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	20.0	0.59	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	25.0	0.67	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	25.0	0.72	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	25.0	0.62	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	27.5	1.05	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	27.5	1.04	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	27.5	1.06	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	32.5	1.16	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	32.5	1.16	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	32.5	1.16	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	35.0	1.01	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	35.0	1.02	(Lürling et al., 2013)
<i>Microcystis aeruginosa</i> PCC7941	Modified WC	80.0	35.0	1.00	(Lürling et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> UTEXLB2061	WC	135.0	25.0	0.50	(Sugimoto et al., 2015)
<i>Microcystis aeruginosa</i> UTEXLB2061	WC	135.0	25.0	0.48	(Sugimoto et al., 2015)
<i>Microcystis aeruginosa</i> UTEXLB2061	WC	135.0	25.0	0.49	(Sugimoto et al., 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	0.00	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	0.01	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	0.01	(Thomas and Litchman, 2015)
¹⁰⁷ <i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	0.00	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.21	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.22	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.21	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.21	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.39	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.41	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.40	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.40	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.35	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.35	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.34	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.35	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	0.68	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	0.66	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	0.67	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	0.67	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	40.0	-0.21	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	40.0	-0.20	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	40.0	-0.23	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	40.0	-0.21	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	-0.04	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	-0.05	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	-0.02	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	15.0	-0.04	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.13	(Thomas and Litchman, 2015)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.12	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.10	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	20.0	0.12	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.26	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.24	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.25	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	25.0	0.25	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.28	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.27	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.26	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	30.0	0.27	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	-0.15	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	-0.16	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	-0.17	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i> bear AC02	WC	100.0	35.0	-0.16	(Thomas and Litchman, 2015)
<i>Microcystis aeruginosa</i>			30.0	0.81	(Chu et al., 2007)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis aeruginosa</i>			30.0	1.09	(Coles and Jones, 2000)
<i>Microcystis aeruginosa</i>			30.0	0.45	(Imai et al., 2009)
<i>Microcystis aeruginosa</i>			32.0	1.60	(Nalewajko and Murphy, 2001)
<i>Microcystis aeruginosa</i>			27.5	0.80	(Nicklisch and Kohl, 1983)
<i>Microcystis aeruginosa</i>			35.0	1.06	(OHKUBO et al., 1991)
<i>Microcystis aeruginosa</i>			25.0	0.19	(Sivonen et al., 1990)
<i>Microcystis aeruginosa</i>			25.0	0.36	(Staeher and Birkeland, 2006)
<i>Microcystis aeruginosa</i>			32.0	0.59	(Watanabe and Oishi, 1985)
<i>Microcystis aeruginosa</i>			32.0	0.81	(van der Westhuizen and Eloff, 1985)
<i>Microcystis aeruginosa</i>		77.3 \pm	30.0 \pm	0.52	(Xiao et al., 2020)
		11.4	18.2		
<i>Microcystis agardhii</i>	Z8	4.0	20.0	0.06	(Salvador et al., 2016)
<i>Microcystis agardhii</i>	Z8	20.0	20.0	0.09	(Salvador et al., 2016)
<i>Microcystis agardhii</i>	Z8	30.0	20.0	0.09	(Salvador et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis ichthyoblabe</i>			35.0	1.32	(Sabour et al., 2009)
<i>Microcystis ma</i>	MLA	60.0	27.0	0.57	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	27.0	0.64	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	27.0	0.50	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	30.0	0.32	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	30.0	0.37	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	30.0	0.26	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	33.0	0.55	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	33.0	0.38	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	33.0	0.73	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	36.0	0.68	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	36.0	0.65	(Mowe et al., 2015)
<i>Microcystis ma</i>	MLA	60.0	36.0	0.62	(Mowe et al., 2015)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	15.0	0.03	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	25.0	0.25	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	35.0	0.26	(Wu et al., 2011)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Microcystis sp. FACHB1271</i>	CT	25.0	15.0	0.04	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	25.0	0.25	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	35.0	0.27	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	15.0	0.03	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	25.0	0.25	(Wu et al., 2011)
<i>Microcystis sp. FACHB1271</i>	CT	25.0	35.0	0.26	(Wu et al., 2011)
<i>Microcystis viridis</i>			25.0	0.17	(OHKUBO et al., 1991)
<i>Microcystis viridis</i>			30.0	0.52	(OHKUBO et al., 1991)
<i>Microcystis wesenbergii</i>			35.0	0.22	(Imai et al., 2009)
<i>Microcystis wesenbergii</i>			35.0	1.50	(OHKUBO et al., 1991)
<i>Microcystis sp.</i>			25.0	0.35	(Konopka and Brock, 1978)
<i>Oscillatoria acuminata</i>	BG11	33.8 ~ 54.1	28.0	0.24	(Tiwari et al., 2001)
<i>Oscillatoria agardhii</i>			30.0	1.12	(Foy et al., 1976)
<i>Oscillatoria agardhii</i>			25.0	0.23	(Sivonen et al., 1990)
<i>Oscillatoria agardhii</i>			35.0	0.64	(TALBOT, 1991)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Oscillatoria agardhii</i>	BG11	33.8 ~ 54.1	28.0	0.09	(Tiwari et al., 2001)
<i>Oscillatoria annae</i>	BG11	33.8 ~ 54.1	28.0	0.17	(Tiwari et al., 2001)
<i>Oscillatoria brevis</i>	BG11	33.8 ~ 54.1	28.0	0.15	(Tiwari et al., 2001)
¹¹³ <i>Oscillatoria cf. chalybea</i>	BG11	40.0	21.5 ~ 28	0.31	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	40.0	19.0	0.12	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	80.0	28.0	0.62	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	73.0	28.0	0.59	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	65.3	28.0	0.50	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	40.4	28.0	0.28	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	31.9	28.0	0.31	(Van Der Ploeg et al., 1995)
<i>Oscillatoria cf. chalybea</i>	BG11	30.3	28.0	0.19	(Van Der Ploeg et al., 1995)
<i>Oscillatoria limosa</i> Agardh ex Gomont	CT	30.0	15.0	0.26	(Cai et al., 2017)
<i>Oscillatoria limosa</i> Agardh ex Gomont	CT	30.0	25.0	0.28	(Cai et al., 2017)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Oscillatoria limosa</i> Agardh ex Gomont	CT	30.0	35.0	0.12	(Cai et al., 2017)
<i>Oscillatoria limosa</i> Agardh ex Gomont	CT	60.0	25.0	0.32	(Cai et al., 2017)
<i>Oscillatoria limosa</i> Agardh ex Gomont	CT	10.0	25.0	0.26	(Cai et al., 2017)
<i>Oscillatoria limosa</i>	BG11	33.8 ~ 54.1	28.0	0.14	(Tiwari et al., 2001)
<i>Oscillatoria mougeotii</i>			30.0	0.56	(Chu et al., 2007)
<i>Oscillatoria priestleyi</i> O-salt	BG20	225.0	20.0	0.29	(Tang et al., 1997)
<i>Oscillatoria proteus</i>	BG11	33.8 ~ 54.1	28.0	0.07	(Tiwari et al., 2001)
<i>Oscillatoria redekei</i>			25.0	1.32	(Foy et al., 1976)
<i>Oscillatoria sancta</i>	BG11	33.8 ~ 54.1	28.0	0.08	(Tiwari et al., 2001)
<i>Oscillatoria simplicissima</i>			28.0	0.22	(Venter et al., 2003)
<i>Oscillatoria simplicissima</i>	BG11	33.8 ~ 54.1	28.0	0.10	(Tiwari et al., 2001)
<i>Oscillatoria</i> sp. E17	BG18	225.0	18.3	0.23	(Tang et al., 1997)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Oscillatoria sp. O-201</i>	BG32	225.0	15.0	0.19	(Tang et al., 1997)
<i>Oscillatoria vizagapatensis</i>	BG11	33.8 ~ 54.1	28.0	0.06	(Tiwari et al., 2001)
<i>Oscillatoria sp.</i>			25.0	1.11	(Coles and Jones, 2000)
<i>Oscillatoria sp.</i>			27.0	0.36	(Novak and Brune, 1985)
<i>Phormidium amoenum Pho012</i>	ASM	100.0	25.0	0.15	(Li et al., 2012)
<i>Phormidium amoenum Pho012</i>	ASM	10.0	25.0	0.07	(Li et al., 2012)
<i>Phormidium amoenum Pho012</i>	ASM	100.0	18.0	0.08	(Li et al., 2012)
<i>Phormidium amoenum Pho012</i>	ASM	10.0	18.0	0.10	(Li et al., 2012)
<i>Phormidium angustissimum</i>	BG11	33.8 ~ 54.1	28.0	0.10	(Tiwari et al., 2001)
<i>Phormidium autumnale O-152</i>	BG28	225.0	24.9	0.15	(Tang et al., 1997)
<i>Phormidium autumnale O-154</i>	BG29	225.0	25.0	0.12	(Tang et al., 1997)
<i>Phormidium bohneri</i>			35.0	1.59	(TALBOT, 1991)
<i>Phormidium bohneri</i>	BG11	33.8 ~ 54.1	28.0	0.21	(Tiwari et al., 2001)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Phormidium foveolarum</i>	BG11	33.8 ~ 54.1	28.0	0.22	(Tiwari et al., 2001)
<i>Phormidium fragile</i>	BG11	33.8 ~ 54.1	28.0	0.18	(Tiwari et al., 2001)
<i>Phormidium murrayi O-099</i>	BG24	225.0	20.0	0.37	(Tang et al., 1997)
<i>Phormidium rubroterricola</i>	BG11	33.8 ~ 54.1	28.0	0.12	(Tiwari et al., 2001)
<i>Phormidium sp. E12</i>	BG17	225.0	24.9	0.24	(Tang et al., 1997)
<i>Phormidium sp. E18</i>	BG19	225.0	20.2	0.29	(Tang et al., 1997)
<i>Phormidium sp. E6a</i>	BG13	225.0	17.7	0.17	(Tang et al., 1997)
<i>Phormidium sp. E7</i>	BG14	225.0	20.8	0.16	(Tang et al., 1997)
<i>Phormidium sp. Ellb</i>	BG16	225.0	18.6	0.28	(Tang et al., 1997)
<i>Phormidium sp. Ella</i>	BG15	225.0	25.5	0.17	(Tang et al., 1997)
<i>Phormidium sp. F5</i>	BG12	225.0	20.1	0.19	(Tang et al., 1997)
<i>Phormidium sp. F9</i>	BG11	225.0	17.9	0.20	(Tang et al., 1997)
<i>Phormidium sp. O-025</i>	BG21	225.0	16.1	0.20	(Tang et al., 1997)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Phormidium</i> sp. O-042	BG22	225.0	15.0	0.31	(Tang et al., 1997)
<i>Phormidium</i> sp. O-043	BG23	225.0	18.9	0.21	(Tang et al., 1997)
<i>Phormidium</i> sp. O-104	BG26	225.0	20.7	0.25	(Tang et al., 1997)
<i>Phormidium</i> sp. O-109	BG25	225.0	19.3	0.20	(Tang et al., 1997)
<i>Phormidium</i> sp. O-120	BG27	225.0	35.0	0.41	(Tang et al., 1997)
<i>Phormidium</i> sp. O-157	BG30	225.0	16.1	0.16	(Tang et al., 1997)
<i>Phormidium</i> sp. O-160	BG31	225.0	20.1	0.13	(Tang et al., 1997)
<i>Phormidium</i> sp. O-202	BG33	225.0	25.0	0.31	(Tang et al., 1997)
<i>Phormidium</i> sp. O-203	BG34	225.0	20.6	0.24	(Tang et al., 1997)
<i>Phormidium</i> sp. O-204	BG35	225.0	19.6	0.26	(Tang et al., 1997)
<i>Phormidium</i> sp. O-210	BG36	225.0	17.0	0.21	(Tang et al., 1997)
<i>Phormidium</i> sp. O-211	BG37	225.0	15.9	0.30	(Tang et al., 1997)
<i>Phormidium</i> sp. Pho689	WC	100.0	25.0	0.09	(Li et al., 2012)
<i>Phormidium</i> sp. Pho689	WC	10.0	25.0	0.14	(Li et al., 2012)
<i>Phormidium</i> sp. Pho689	WC	100.0	18.0	0.08	(Li et al., 2012)
<i>Phormidium</i> sp. Pho689	WC	10.0	18.0	0.07	(Li et al., 2012)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Planktothricoides raciborskii</i>	BG11			0.39	This study
<i>Planktothricoides raciborskii</i>	BG11			1.96	This study
<i>Planktothricoides raciborskii</i>	BG11			0.39	This study
<i>Planktothricoides raciborskii</i>	BG11			0.08	This study
<i>Planktothricoides raciborskii</i>	BG11			0.27	This study
<i>Planktothricoides raciborskii</i>	BG11			0.27	This study
<i>Planktothrix agardhii</i>	Modified MIV	145.0	20.0	0.38	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	92.0	20.0	0.34	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	145.0	20.0	0.42	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	160.0	20.0	0.62	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	61.0	20.0	0.51	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	308.0	20.0	0.42	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	64.0	20.0	0.66	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	237.0	20.0	0.56	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	309.0	20.0	0.58	(Nicklisch, 1998)
<i>Planktothrix agardhii</i>	Modified MIV	467.0	20.0	0.29	(Nicklisch, 1998)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Planktothrix agardhii</i>	Modified MIII		28.0	0.74	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		24.0	0.72	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		20.0	0.58	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		16.0	0.43	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		12.0	0.23	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		8.0	0.13	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		20.0	0.56	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		20.0	0.57	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i>	Modified MIII		20.0	0.44	(Nicklisch et al., 2007)
<i>Planktothrix agardhii</i> CYA116			20.0	0.50	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA116			25.0	0.71	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA116			22.5	0.82	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA116			27.5	NA	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA116			25.0	0.70	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA116			30.0	0.40	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA126			20.0	0.43	(Lürling et al., 2013)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Planktothrix agardhii</i> CYA126			25.0	0.60	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA126			22.5	NA	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA126			27.5	0.58	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA126			25.0	NA	(Lürling et al., 2013)
<i>Planktothrix agardhii</i> CYA126			30.0	NA	(Lürling et al., 2013)
<i>Planktothrix</i> sp.	BG11	5.0	25.0	0.02	(Jia et al., 2019)
<i>Planktothrix</i> sp.	BG11	17.0	25.0	0.16	(Jia et al., 2019)
<i>Planktothrix</i> sp.	BG11	36.0	25.0	0.20	(Jia et al., 2019)
<i>Planktothrix</i> sp.	BG11	85.0	25.0	0.16	(Jia et al., 2019)
<i>Planktothrix</i> sp.	BG11	250.0	25.0	0.18	(Jia et al., 2019)
<i>Planktothrix</i> sp.	BG11	4.4	25.0	0.00	(Jia et al., 2019)
<i>Plectonema battersii</i>	BG11	33.8 ~ 54.1	28.0	0.18	(Tiwari et al., 2001)
<i>Plectonema boryanurn</i> UTEX-482				1.44	(Prakash et al., 1999)
<i>Plectonema boryanurn</i> UTEX-482				1.40	(Prakash et al., 1999)
<i>Plectonema boryanurn</i> UTEX-485				0.29	(Miskiewicz et al., 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Plectonema boryanurn UTEX-485</i>				2.05	(Miskiewicz et al., 2000)
<i>Plectonema boryanurn UTEX-485</i>				0.39	(Miskiewicz et al., 2000)
<i>Plectonema boryanurn UTEX-485</i>				2.15	(Miskiewicz et al., 2000)
<i>Plectonema golenkilianum</i>	BG11	33.8 ~ 54.1	28.0	0.07	(Tiwari et al., 2001)
<i>Plectonema nostocorum</i>	BG11	33.8 ~ 54.1	28.0	0.12	(Tiwari et al., 2001)
<i>Plectonema yellowstonense</i>	BG11	33.8 ~ 54.1	28.0	0.14	(Tiwari et al., 2001)
<i>Pseudanabaena frigidum</i>	BG11	33.8 ~ 54.1	28.0	0.26	(Tiwari et al., 2001)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	25.0	25.0	0.25	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	10.0	25.0	0.11	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	40.0	25.0	0.23	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	55.0	25.0	0.19	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	70.0	25.0	0.17	(Zhang et al., 2016)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena sp. FACHB1277</i>	BG11	85.0	25.0	0.15	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	25.0	10.0	0.08	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	25.0	15.0	0.19	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	25.0	20.0	0.24	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	25.0	30.0	0.24	(Zhang et al., 2016)
<i>Pseudanabaena sp. FACHB1277</i>	BG11	25.0	35.0	0.20	(Zhang et al., 2016)
<i>Pseudanabaena sp. dqh15</i>	CT	10.0	25.0	0.08	(Wang and Li, 2015)
<i>Pseudanabaena sp. dqh15</i>	CT	30.0	25.0	0.09	(Wang and Li, 2015)
<i>Pseudanabaena sp. dqh15</i>	CT	60.0	25.0	0.04	(Wang and Li, 2015)
<i>Pseudanabaena sp. dqh15</i>	CT	30.0	10.0	0.18	(Wang and Li, 2015)
<i>Pseudanabaena sp. dqh15</i>	CT	30.0	35.0	0.07	(Wang and Li, 2015)
<i>Pseudanabaena sp.</i>	TN: 247 mg/L,TP: 7.12 mg/L	71.0	15.0	0.13	(Gao et al., 2018)
<i>Pseudanabaena sp.</i>	TN: 247 mg/L,TP: 7.12 mg/L	71.0	20.0	0.17	(Gao et al., 2018)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.19	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	30.0	0.16	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	35.0	0.13	(Gao et al., 2018)
123 <i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	40.0	-0.02	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	0.0	25.0	-0.13	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	9.0	25.0	0.18	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	18.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	27.0	25.0	0.23	(Gao et al., 2018)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	36.0	25.0	0.22	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	45.0	25.0	0.22	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.22	(Gao et al., 2018)
124 <i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	142.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	216.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:0 mg/L,TP: 7.12 mg/L	71.0	25.0	-0.11	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:2.4mg/L,TP: 7.12 mg/L	71.0	25.0	-0.01	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:4.8mg/L,TP: 7.12 mg/L	71.0	25.0	0.04	(Gao et al., 2018)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN:9.6mg/L,TP: 7.12 mg/L	71.0	25.0	0.09	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:19.2mg/L,TP: 7.12 mg/L	71.0	25.0	0.17	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:38.4mg/L,TP: 7.12 mg/L	71.0	25.0	0.19	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:76.8mg/L,TP: 7.12 mg/L	71.0	25.0	0.20	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:153.6mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN:230.4mg/L,TP: 7.12 mg/L	71.0	25.0	0.22	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP:0mg/L	71.0	25.0	0.06	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP:0.04mg/L	71.0	25.0	0.22	(Gao et al., 2018)

125

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 0.16 mg/L	71.0	25.0	0.25	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 0.32 mg/L	71.0	25.0	0.27	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 0.64 mg/L	71.0	25.0	0.28	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 1.28 mg/L	71.0	25.0	0.28	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 5.12 mg/L	71.0	25.0	0.24	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 10.24 mg/L	71.0	25.0	0.23	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 102.4 mg/L	71.0	25.0	0.05	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L, TP: 7.12 mg/L	71.0	25.0	0.00	(Gao et al., 2018)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.24	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.23	(Gao et al., 2018)
127 <i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.24	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.22	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.00	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.00	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.25	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.25	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.22	(Gao et al., 2018)
128 <i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.20	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.20	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.20	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)
<i>Pseudanabaena</i> sp.	TN: 247 mg/L,TP: 7.12 mg/L	71.0	25.0	0.21	(Gao et al., 2018)
<i>Selenastrum capricornutum</i>			30.0	0.61	(Fujimoto et al., 1994)
<i>Selenastrum capricornutum</i>			21.0	1.08	(Novak and Brune, 1985)
<i>Selenastrum minutum</i>			35.0	1.73	(Bouarab et al., 2002)
<i>Synechococcus elongatus</i> PCC6301			20.0	0.30	(Lürling et al., 2013)
<i>Synechococcus elongatus</i> PCC6301			25.0	0.67	(Lürling et al., 2013)
<i>Synechococcus elongatus</i> PCC6301			22.5	0.64	(Lürling et al., 2013)
<i>Synechococcus elongatus</i> PCC6301			27.5	0.72	(Lürling et al., 2013)
<i>Synechococcus elongatus</i> PCC6301			25.0	0.82	(Lürling et al., 2013)
<i>Synechococcus elongatus</i> PCC6301			30.0	0.91	(Lürling et al., 2013)
<i>Synechococcus</i> sp. CCMP1768		43.0		0.23	(Glibert et al., 2009)
<i>Synechococcus</i> sp. CCMP1768		43.0		0.49	(Glibert et al., 2009)
<i>Synechococcus</i> sp. strain PCC6301				0.03	(Lepp and Schmidt, 2004)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Synechococcus sp. strain PCC6301</i>				0.48	(Lepp and Schmidt, 2004)
<i>Synechococcus sp. strain PCC6301</i>				3.40	(Lepp and Schmidt, 2004)
<i>Synechococcus sp. strain PCC6301</i>				1.37	(Lepp and Schmidt, 2004)
<i>Synechococcus sp. strain PCC6301</i>				0.19	(Lepp and Schmidt, 2004)
<i>Synechococcus spp.</i>	Nitrate:1.7 μM		25.2	2.16	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.1 μM		26.5	3.12	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.1 μM		26.6	-0.24	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0 μM		28.2	0.24	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.4 μM		28.6	1.44	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.4 μM		27.4	0.96	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:4.6 μM		24.8	9.36	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:37.2 μM		22.5	0.72	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:29.5 μM		24.7	2.40	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.3 μM		25.3	-1.92	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.6 μM		26.4	4.08	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.4 μM		26.1	2.16	(Gong and Tsai, 2019)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Synechococcus spp.</i>	Nitrate:0.3 μM		25.9	-0.24	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.2 μM		26.1	1.68	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>	Nitrate:0.1 μM		29.9	1.44	(Gong and Tsai, 2019)
<i>Synechococcus spp.</i>				0.70	(Liu et al., 1998)
<i>Synechococcus spp.</i>				1.33	(Liu et al., 1998)
<i>Synechococcus spp.</i>				0.44	(Liu et al., 1998)
<i>Synechococcus spp.</i>				1.35	(Liu et al., 1998)
<i>Synechococcus spp.</i>				1.49	(Liu et al., 1998)
<i>Synechococcus spp.</i>	NO ₃ :2.3 μM ; PO ₄ :0.4 μM		28.5	0.60	(Tsai and Mukhanov, 2021)
<i>Synechococcus spp.</i>	NO ₃ :19.4 μM ; PO ₄ :1.9 μM			0.66	(Tsai and Mukhanov, 2021)
<i>Synechococcus spp.</i>	NO ₃ :1.8 μM ; PO ₄ :0.5 μM		30.0	0.62	(Tsai and Mukhanov, 2021)
<i>Synechococcus spp.</i>	NO ₃ :18.6 μM ; PO ₄ :1.8 μM			0.65	(Tsai and Mukhanov, 2021)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Synechococcus spp.</i>	NO ₃ :3.1 μM ; PO ₄ :0.8 μM		29.0	0.59	(Tsai and Mukhanov, 2021)
<i>Synechococcus spp.</i>	NO ₃ :16.2 μM ; PO ₄ :1.5 μM			0.64	(Tsai and Mukhanov, 2021)
<i>Synechococcus spp.</i>	NO ₃ :3.5 μM ; PO ₄ :0.7 μM		28.5	0.54	(Tsai and Mukhanov, 2021)
¹³² <i>Synechococcus spp.</i>	NO ₃ :18.9 μM ; PO ₄ :1.6 μM			0.56	(Tsai and Mukhanov, 2021)
<i>Synechococcus spp.</i>				-0.25	(Heng et al., 2017)
<i>Synechococcus spp.</i>				0.98	(Heng et al., 2017)
<i>Synechococcus spp.</i>				0.29	(Heng et al., 2017)
<i>Synechococcus spp.</i>				0.82	(Heng et al., 2017)
<i>Synechococcus sp.</i>			20.0	0.98	(Malinsky-Rushansky, 2002)
<i>Synechococcus sp.</i>			28.0	1.19	(Malinsky-Rushansky, 2002)
<i>Synechococcus sp.</i>	N:2.54 μM N d ⁻¹ ; P:1/20N			1.17	(Agawin et al., 2000)

Table 10 The summary of the main cyanobacteria specific growth rates under different culture conditions

Genus/Strain	Nutrient cond.	Light (μE)	Temp. ($^{\circ}\text{C}$)	Growth rate (d^{-1})	Ref.
<i>Synechococcus</i> sp.	0~10.18 μM N d ⁻¹ ; P:1/20N			5.00	(Agawin et al., 2000)
<i>Synechococcus</i> sp.	0.25 μM N d ⁻¹ ; P:1/20N			3.05	(Agawin et al., 2000)

36 References

- 37 Agawin, N., Duarte, C., Agustí, S., 2000. Response of mediterranean synechococcus growth and loss rates to experimental
38 nutrient inputs. *Marine Ecology Progress Series* 206, 97–106. URL: <https://doi.org/10.3354/meps206097>, doi:10.
39 [3354/meps206097](https://doi.org/10.3354/meps206097).
- 40 Bañares-España, E., Kromkamp, J.C., López-Rodas, V., Costas, E., Flores-Moya, A., 2012. Photoacclimation of cultured
41 strains of the cyanobacterium *Microcystis aeruginosa* to high-light and low-light conditions. *FEMS Microbiology*
42 *Ecology* 83, 700–710. URL: <https://doi.org/10.1111/1574-6941.12025>, doi:10.1111/1574-6941.12025.
- 43 Bonilla, S., Aubriot, L., Soares, M.C.S., González-piana, M., Fabre, A., Huszar, V.L., Lürling, M., Antoniadis, D., Padiśák,
44 J., Kruk, C., 2012. What drives the distribution of the bloom-forming cyanobacteria *Planktothrix agardhii* and
45 *Cylindrospermopsis Raciborskii*? *Fems Microbiology Ecology* 79, 594–607. URL: [http://dx.doi.org/10.1111/j.1574-](http://dx.doi.org/10.1111/j.1574-6941.2011.01242.x)
46 [6941.2011.01242.x](http://dx.doi.org/10.1111/j.1574-6941.2011.01242.x), doi:10.1111/j.1574-6941.2011.01242.x.
- 47 Bouarab, L., Loudiki, M., Dauta, A., 2002. Croissance en autotrophie et en mixotrophie de la microalgue *Micractinium*
48 *pusillum* fres. isolée d'un lagunage naturel : influence de la lumière et de la température. *Journal of Water Science*
49 15, 73–86. doi:<https://doi.org/10.7202/705437ar>.
- 50 Briand, E., Bormans, M., Quiblier, C., Salençon, M.J., Humbert, J.F., 2012. Evidence of the cost of the production of mi-
51 crocystins by *microcystis aeruginosa* under differing light and nitrate environmental conditions. *PLoS ONE* 7, e29981.
52 URL: <https://doi.org/10.1371/journal.pone.0029981>, doi:10.1371/journal.pone.0029981.
- 53 Briand, J.F., Leboulanger, C., Humbert, J.F., Bernard, C., Dufour, P., 2004. CYLINDROSPERMOPSIS RACIBORSKII
54 (CYANOBACTERIA) INVASION AT MID-LATITUDES: SELECTION, WIDE PHYSIOLOGICAL TOLERANCE, ORGLOBALWARM-
55 ING?1. *Journal of Phycology* 40, 231–238. URL: <https://doi.org/10.1111/j.1529-8817.2004.03118.x>, doi:10.1111/
56 [j.1529-8817.2004.03118.x](https://doi.org/10.1111/j.1529-8817.2004.03118.x).
- 57 Cai, F., Yu, G., Zhang, K., Chen, Y., Li, Q., Yang, Y., Xie, J., Wang, Y., Li, R., 2017. Geosmin production and polyphasic
58 characterization of *Oscillatoria limosa* agardh ex gomont isolated from the open canal of a large drinking water system
59 in tianjin city, china. *Harmful Algae* 69, 28–37. URL: <https://doi.org/10.1016/j.hal.2017.09.006>, doi:10.1016/j.
60 [hal.2017.09.006](https://doi.org/10.1016/j.hal.2017.09.006).
- 61 Chu, Z., Jin, X., Iwami, N., Inamori, Y., 2007. The effect of temperature on growth characteristics and competitions of
62 *Microcystis aeruginosa* and *Oscillatoria mougeotii* in a shallow, eutrophic lake simulator system, in: *Eutrophication*
63 *of Shallow Lakes With Special Reference to Lake Taihu, China*. Springer Netherlands. volume 194, pp. 217–223. URL:
64 http://dx.doi.org/10.1007/978-1-4020-6158-5_24, doi:10.1007/978-1-4020-6158-5_24.
- 65 Coles, J.F., Jones, R.C., 2000. Effect of temperature on photosynthesis-light response and growth of four phytoplankton
66 species isolated from a tidal freshwater river. *Journal of Phycology* 36, 7–16. URL: [https://doi.org/10.1046/j.1529-](https://doi.org/10.1046/j.1529-8817.2000.98219.x)
67 [8817.2000.98219.x](https://doi.org/10.1046/j.1529-8817.2000.98219.x), doi:10.1046/j.1529-8817.2000.98219.x.
- 68 Elmetri, I., Bell, P., 2004. Effects of phosphorus on the growth and nitrogen fixation rates of *Lyngbya majuscula*:
69 implications for management in Moreton Bay, Queensland. *Marine Ecology Progress Series* 281, 27–35. URL:
70 <https://doi.org/10.3354/meps281027>, doi:10.3354/meps281027.

71 Foy, R., Gibson, C., Smith, R., 1976. The influence of daylength, light intensity and temperature on the growth
72 rates of planktonic blue-green algae. *British Phycological Journal* 11, 151–163. URL: [https://doi.org/10.1080/](https://doi.org/10.1080/00071617600650181)
73 [00071617600650181](https://doi.org/10.1080/00071617600650181), doi:10.1080/00071617600650181.

74 Fujimoto, N., Inamori, Y., Sugiura, N., Sudo, R., 1994. Effects of temperature change on algal growth. *Environmental*
75 *Technology* 15, 497–500. URL: <https://doi.org/10.1080/09593339409385455>, doi:10.1080/09593339409385455.

76 Gao, J., Zhu, J., Wang, M., Dong, W., 2018. Dominance and growth factors of *Pseudanabaena* sp. in drinking water
77 source reservoirs, southern china. *Sustainability* 10, 3936. URL: [https://doi.org/10.3390/](https://doi.org/10.3390/su10113936)
78 [su10113936](https://doi.org/10.3390/su10113936), doi:10.3390/
79 [su10113936](https://doi.org/10.3390/su10113936).

79 Giraldo-Silva, A., Nelson, C., Penfold, C., Barger, N.N., Garcia-Pichel, F., 2019. Effect of preconditioning to the soil en-
80 vironment on the performance of 20 cyanobacterial strains used as inoculum for biocrust restoration. *Restoration*
81 *Ecology* 28. URL: <https://doi.org/10.1111/rec.13048>, doi:10.1111/rec.13048.

82 Glibert, P., Burkholder, J., Kana, T., Alexander, J., Skelton, H., Shilling, C., 2009. Grazing by *karenia brevis* on *synechococcus*
83 enhances its growth rate and may help to sustain blooms. *Aquatic Microbial Ecology* 55, 17–30. URL: [https://doi.](https://doi.org/10.3354/ame01279)
84 [org/10.3354/ame01279](https://doi.org/10.3354/ame01279), doi:10.3354/ame01279.

85 Gong, G.C., Tsai, A.Y., 2019. Reduced daytime net growth rate of *synechococcus* spp. in the east china sea in summer
86 estimated using a dilution approach. *Estuarine, Coastal and Shelf Science* 219, 90–96. URL: [https://doi.org/10.1016/](https://doi.org/10.1016/j.ecss.2019.01.029)
87 [j.ecss.2019.01.029](https://doi.org/10.1016/j.ecss.2019.01.029), doi:10.1016/j.ecss.2019.01.029.

88 Heng, P.L., Lim, J.H., Lee, C.W., 2017. *Synechococcus* production and grazing loss rates in nearshore tropical waters.
89 *Environmental Monitoring and Assessment* 189. URL: <https://doi.org/10.1007/s10661-017-5838-1>, doi:10.1007/
90 [s10661-017-5838-1](https://doi.org/10.1007/s10661-017-5838-1).

91 Hesse, K., Dittmann, E., Bärner, T., 2001. Consequences of impaired microcystin production for light-dependent growth
92 and pigmentation of *microcystis aeruginosa* PCC 7806. *FEMS Microbiology Ecology* 37, 39–43. URL: [https://doi.org/](https://doi.org/10.1111/j.1574-6941.2001.tb00851.x)
93 [10.1111/j.1574-6941.2001.tb00851.x](https://doi.org/10.1111/j.1574-6941.2001.tb00851.x), doi:10.1111/j.1574-6941.2001.tb00851.x.

94 Imai, H., Chang, K.h., Kusaba, M., Nakano, S.i., 2009. Temperature-dependent dominance of *Microcys-*
95 *tis* (cyanophyceae) species: *M. aeruginosa* and *M. wesenbergii*. *Journal of Plankton Research* 31,
96 171–178. URL: <http://plankt.oxfordjournals.org/content/31/2/171.abstract>, doi:10.1093/plankt/fbn110,
97 [arXiv:http://plankt.oxfordjournals.org/content/31/2/171.full.pdf+html](http://plankt.oxfordjournals.org/content/31/2/171.full.pdf+html).

98 Jia, Z., Su, M., Liu, T., Guo, Q., Wang, Q., Burch, M., Yu, J., Yang, M., 2019. Light as a possible regulator of MIB-producing
99 *Planktothrix* in source water reservoir, mechanism and *in-situ* verification. *Harmful Algae* 88, 101658. URL: [http:](http://www.sciencedirect.com/science/article/pii/S1568988319301313)
100 [//www.sciencedirect.com/science/article/pii/S1568988319301313](http://www.sciencedirect.com/science/article/pii/S1568988319301313), doi:10.1016/j.hal.2019.101658.

101 Karsten, U., 1996. Growth and organic osmolytes of geographically different isolates of mi-
102 crocoleus chthonoplastes (cyanobacteria) from benthic microbial mats: response to salinity
103 change1. *Journal of Phycology* 32, 501–506. URL: [https://onlinelibrary.wiley.com/doi/abs/10.](https://onlinelibrary.wiley.com/doi/abs/10.1111/j.0022-3646.1996.00501.x)
104 [1111/j.0022-3646.1996.00501.x](https://onlinelibrary.wiley.com/doi/abs/10.1111/j.0022-3646.1996.00501.x),
105 [doi:https://doi.org/10.1111/j.0022-3646.1996.00501.x](https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.0022-3646.1996.00501.x),
[arXiv:https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.0022-3646.1996.00501.x](https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.0022-3646.1996.00501.x).

- 106 Konopka, A., Brock, T.D., 1978. Effect of temperature on blue-green algae (cyanobacteria) in Lake Mendota. Applied
107 and Environmental Microbiology 36, 572–576. URL: <https://doi.org/10.1128/aem.36.4.572-576.1978>, doi:10.1128/
108 [aem.36.4.572-576.1978](https://doi.org/10.1128/aem.36.4.572-576.1978).
- 109 Kratz, W.A., Myers, J., 1955. NUTRITION AND GROWTH OF SEVERAL BLUE-GREEN ALGAE. American Journal of
110 Botany 42, 282–287. URL: <https://doi.org/10.1002/j.1537-2197.1955.tb11120.x>, doi:10.1002/j.1537-2197.
111 [1955.tb11120.x](https://doi.org/10.1002/j.1537-2197.1955.tb11120.x).
- 112 Lababpour, A., Kaviani, M., 2016. Isolation and submerged culture biomass production of the arid land cyanobacteria
113 microcoleus spp., an investigation on its utilization for biological soil crust restoration. Environmental Earth Sciences
114 75. URL: <https://doi.org/10.1007/s12665-016-6313-y>, doi:10.1007/s12665-016-6313-y.
- 115 Lehtimäki, J., Moisander, P., Sivonen, K., Kononen, K., 1997. Growth, nitrogen fixation, and nodularin production by two
116 Baltic Sea cyanobacteria. Applied and Environmental Microbiology 63, 1647–1656. URL: <https://doi.org/10.1128/aem.63.5.1647-1656.1997>,
117 doi:10.1128/aem.63.5.1647-1656.1997.
- 118 Lepp, P., Schmidt, T., 2004. Changes in synechococcus population size and cellular ribosomal RNA content in response
119 to predation and nutrient limitation. Microbial Ecology 48, 1–9. URL: <https://doi.org/10.1007/s00248-003-1041-8>,
120 doi:10.1007/s00248-003-1041-8.
- 121 Li, M., Nkrumah, P.N., Xiao, M., 2014. Biochemical composition of microcystis aeruginosa re-
122 lated to specific growth rate: insight into the effects of abiotic factors. Inland Waters 4, 357–
123 362. URL: <https://www.tandfonline.com/doi/abs/10.5268/IW-4.4.710>, doi:10.5268/IW-4.4.710,
124 arXiv:<https://www.tandfonline.com/doi/pdf/10.5268/IW-4.4.710>.
- 125 Li, Z., Hobson, P., An, W., Burch, M.D., House, J., Yang, M., 2012. Earthy odor compounds production and loss in three
126 cyanobacterial cultures. Water Research 46, 5165–5173. URL: <http://www.sciencedirect.com/science/article/pii/S0043135412004046>, doi:10.1016/j.watres.2012.06.008.
127
- 128 Liu, H., Campbell, L., Landry, M., Nolla, H., Brown, S., Constantinou, J., 1998. Prochlorococcus and synechococcus
129 growth rates and contributions to production in the arabian sea during the 1995 southwest and northeast monsoons.
130 Deep Sea Research Part II: Topical Studies in Oceanography 45, 2327–2352. URL: [https://doi.org/10.1016/S0967-0645\(98\)00073-3](https://doi.org/10.1016/S0967-0645(98)00073-3),
131 doi:10.1016/S0967-0645(98)00073-3.
- 132 Lürling, M., Eshetu, F., Faassen, E.J., Kosten, S., Huszar, V.L.M., 2013. Comparison of cyanobacterial and green al-
133 gal growth rates at different temperatures. Freshwater Biology 58, 552–559. doi:10.1111/j.1365-2427.2012.
134 [02866.x](https://doi.org/10.1111/j.1365-2427.2012.02866.x).
- 135 Malinsky-Rushansky, N., 2002. Physiological characteristics of picophytoplankton, isolated from lake kinneret: responses
136 to light and temperature. Journal of Plankton Research 24, 1173–1183. URL: <https://doi.org/10.1093/plankt/24.11.1173>,
137 doi:10.1093/plankt/24.11.1173.
- 138 Marinho, M.M., Souza, M.B.G., Lürling, M., 2013. Light and phosphate competition between cylindrospermopsis raci-
139 borskii and microcystis aeruginosa is strain dependent. Microbial Ecology 66, 479–488. URL: <https://doi.org/10.1007/s00248-013-0232-1>,
140 doi:10.1007/s00248-013-0232-1.

141 Mehnert, G., Leunert, F., Cires, S., Johnk, K.D., Rucker, J., Nixdorf, B., Wiedner, C., 2010. Competitiveness of invasive
142 and native cyanobacteria from temperate freshwaters under various light and temperature conditions. *Journal of*
143 *Plankton Research* 32, 1009–1021. URL: <https://doi.org/10.1093/plankt/fbq033>, doi:10.1093/plankt/fbq033.

144 e. Mello, M.M., Soares, M.C.S., Roland, F., Lurling, M., 2012. Growth inhibition and colony formation in the cyanobac-
145 terium *Microcystis aeruginosa* induced by the cyanobacterium *Cylindrospermopsis raciborskii*. *Journal of Plankton*
146 *Research* 34, 987–994. URL: <https://doi.org/10.1093/plankt/fbs056>, doi:10.1093/plankt/fbs056.

147 Miskiewicz, E., Ivanov, A.G., Williams, J.P., Khan, M.U., Falk, S., Huner, N.P., 2000. Photosynthetic acclimation of the
148 filamentous cyanobacterium, *Plectonema boryanum* UTEX 485, to temperature and light. *Plant and Cell Physiology*
149 41, 767–775. URL: <https://doi.org/10.1093/pcp/41.6.767>, doi:10.1093/pcp/41.6.767.

150 Mowe, M.A.D., Abbas, F., Porojan, C., Mitrovic, S.M., Lim, R.P., Furey, A., Yeo, D.C.J., 2015. Roles of nitrogen and
151 phosphorus in growth responses and toxin production (using LC-MS/MS) of tropical *Microcystis ichthyoblabe* and
152 *M. flos-aquae*. *Journal of Applied Phycology* 28, 1543–1552. URL: <https://doi.org/10.1007/s10811-015-0688-0>,
153 doi:10.1007/s10811-015-0688-0.

154 Nalewajko, C., Murphy, T.P., 2001. Effects of temperature, and availability of nitrogen and phosphorus on the abundance
155 of *Anabaena* and *Microcystis* in Lake Biwa, Japan: An experimental approach. *Limnology* 2, 45–48. URL: [http://dx.
156 doi.org/10.1007/s102010170015](http://dx.doi.org/10.1007/s102010170015), doi:10.1007/s102010170015.

157 Nicklisch, A., 1992. The interaction of irradiance and temperature on the growth rate of *Limnithrix redekei*
158 and its mathematical description. *Algological Studies/Archiv für Hydrobiologie, Supplement Volumes* 63, 1–
159 18. URL: [https://www.schweizerbart.de/papers/archiv_algolstud/detail/63/66818/The_interaction_of_irradiance_
160 and_temperature_on_the_growth_rate_of_Limnithrix_redekei_and_its_mathematical_description](https://www.schweizerbart.de/papers/archiv_algolstud/detail/63/66818/The_interaction_of_irradiance_and_temperature_on_the_growth_rate_of_Limnithrix_redekei_and_its_mathematical_description).

161 Nicklisch, A., 1998. Growth and light absorption of some planktonic cyanobacteria, diatoms and chlorophyceae under
162 simulated natural light fluctuations. *Journal of Plankton Research* 20, 105–119. URL: [https://doi.org/10.1093/plankt/
163 20.1.105](https://doi.org/10.1093/plankt/20.1.105), doi:10.1093/plankt/20.1.105.

164 Nicklisch, A., 1999. Competition between the cyanobacterium *Limnithrix redekei* and some spring
165 species of diatoms under p-limitation. *International Review of Hydrobiology* 84, 233–241. URL: <https://onlinelibrary.wiley.com/doi/abs/10.1002/iroh.199900024>, doi:<https://doi.org/10.1002/iroh.199900024>,
166 [arXiv:https://onlinelibrary.wiley.com/doi/pdf/10.1002/iroh.199900024](https://onlinelibrary.wiley.com/doi/pdf/10.1002/iroh.199900024).

167 Nicklisch, A., Kohl, J.G., 1983. Growth kinetics of *Microcystis aeruginosa* (KÜTZ)KÜTZ as a basis for modelling its
168 population dynamics. *Internationale Revue der gesamten Hydrobiologie und Hydrographie* 68, 317–326. URL:
169 <https://doi.org/10.1002/iroh.19830680304>, doi:10.1002/iroh.19830680304.

170 Nicklisch, A., Shatwell, T., Kohler, J., 2007. Analysis and modelling of the interactive effects of temperature and light
171 on phytoplankton growth and relevance for the spring bloom. *Journal of Plankton Research* 30, 75–91. URL: <https://doi.org/10.1093/plankt/fbm099>, doi:10.1093/plankt/fbm099.

172 Novak, J.T., Brune, D.E., 1985. Inorganic carbon limited growth kinetics of some freshwater algae. *Water Research* 19,
173 215–225. URL: [https://doi.org/10.1016/0043-1354\(85\)90203-9](https://doi.org/10.1016/0043-1354(85)90203-9), doi:10.1016/0043-1354(85)90203-9.

- 176 OHKUBO, N., YAGI, O., OKADA, M., 1991. Effects of temperature and illumination on the growth of blue-green alga
177 *Microcystis viridis*. Japanese Journal of Limnology (Rikusuigaku Zasshi) 52, 255–261. URL: [https://doi.org/10.3739/](https://doi.org/10.3739/rikusui.52.255)
178 [rikusui.52.255](https://doi.org/10.3739/rikusui.52.255), doi:10.3739/rikusui.52.255.
- 179 Prakash, A., Margaritis, A., Saunders, R.C., Vijayan, S., 1999. High concentrations ammonia removal by the cyanobac-
180 terium *Plectonema boryanum* in a photobioreactor system. The Canadian Journal of Chemical Engineering 77, 99–
181 106. URL: <https://doi.org/10.1002/cjce.5450770116>, doi:10.1002/cjce.5450770116.
- 182 Rapala, J., Sivonen, K., Luukkainen, R., Niemelä, S.I., 1993. Anatoxin-a concentration in *Anabaena* and *Aphanizomenon*
183 under different environmental conditions and comparison of growth by toxic and non-toxic *Anabaena*-strains — a
184 laboratory study. Journal of Applied Phycology 5, 581–591. URL: [https://doi.org/10.1007/](https://doi.org/10.1007/bf02184637)
185 [bf02184637](https://doi.org/10.1007/bf02184637), doi:10.1007/
- 186 Rapala, J., Sivonen, K., Lyra, C., Niemelä, S.I., 1997. Variation of microcystins, cyanobacterial hepatotoxins, in *Anabaena*
187 spp. as a function of growth stimuli. Applied and Environmental Microbiology 63, 2206–12. URL: [http://aem.asm.](http://aem.asm.org/content/63/6/2206.abstract)
188 [org/content/63/6/2206.abstract](http://aem.asm.org/content/63/6/2206.abstract).
- 189 Sabour, B., Loudiki, M., Vasconcelos, V., 2009. Growth responses of *Microcystis ichthyoblabe* Kützinger and *Anabaena*
190 *aphanizomenoides* Forti (cyanobacteria) under different nitrogen and phosphorus conditions. Chemistry and Ecology
191 25, 337–344. URL: <https://doi.org/10.1080/02757540903193130>, doi:10.1080/02757540903193130.
- 192 Saker, M.L., Griffiths, D.J., 2000. The effect of temperature on growth and cylindrospermopsin content of seven isolates
193 of *Cylindrospermopsis raciborskii* (Nostocales, Cyanophyceae) from water bodies in northern Australia. Phycologia 39,
194 349–354. URL: <https://doi.org/10.2216/i0031-8884-39-4-349.1>, doi:10.2216/i0031-8884-39-4-349.1.
- 195 Saker, M.L., Thomas, A.D., Norton, J.H., 1999. Cattle mortality attributed to the toxic cyanobacterium *Cylindrospermopsis*
196 *raciborskii* in an outback region of north Queensland. Environmental Toxicology 14, 179–182.
197 URL: [https://doi.org/10.1002/\(sici\)1522-7278\(199902\)14:1<179::aid-tox23>3.0.co;](https://doi.org/10.1002/(sici)1522-7278(199902)14:1<179::aid-tox23>3.0.co;2-g)
198 [2-g](https://doi.org/10.1002/(sici)1522-7278(199902)14:1<179::aid-tox23>3.0.co;2-g), doi:10.1002/(sici)1522-7278(199902)14:1<179::aid-tox23>3.0.co;2-g.
- 199 Salvador, D., Churro, C., Valério, E., 2016. Evaluating the influence of light intensity in *mcyA* gene expression and mi-
200 crocystin production in toxic strains of *Planktothrix agardhii* and *Microcystis aeruginosa*. Journal of Microbiological
201 Methods 123, 4–12. URL: <https://doi.org/10.1016/j.mimet.2016.02.002>, doi:10.1016/j.mimet.2016.02.002.
- 202 Seki, H., Ozawa, H., Ichimura, S., 1981. Temperature dependence of filament length of *Anabaena spiroides* Klebahn var.
203 *crassa* Lemm. Hydrobiologia 83, 419–423. URL: <https://doi.org/10.1007/bf02187039>, doi:10.1007/bf02187039.
- 204 Shatwell, T., Nicklisch, A., Köhler, J., 2012. Temperature and photoperiod effects on phytoplankton growing under sim-
205 ulated mixed layer light fluctuations. Limnology and Oceanography 57, 541–553. URL: [https://doi.org/10.4319/lo.](https://doi.org/10.4319/lo.2012.57.2.0541)
206 [2012.57.2.0541](https://doi.org/10.4319/lo.2012.57.2.0541), doi:10.4319/lo.2012.57.2.0541.
- 207 Shen, H., Song, L., 2007. Comparative studies on physiological responses to phosphorus in two phenotypes of bloom-
208 forming *Microcystis*. Hydrobiologia 592, 475–486. URL: <https://doi.org/10.1007/s10750-007-0794-3>, doi:10.1007/
- 209 [s10750-007-0794-3](https://doi.org/10.1007/s10750-007-0794-3).
- 210 Shen, Q., Shimizu, K., Miao, H., Tsukino, S., Utsumi, M., Lei, Z., Zhang, Z., Nishimura, O., Asada, Y., Fujimoto, N., Takanashi,

211 H., Akiba, M., 2020. Effects of elevated nitrogen on the growth and geosmin productivity of *dolichospermum smithii*.
 212 Environmental Science and Pollution Research 28, 177–184. URL: <https://doi.org/10.1007/s11356-020-10429-4>,
 213 doi:10.1007/s11356-020-10429-4.

214 Sivonen, K., Niemelä, S.I., Niemi, R.M., Lepistö, L., Luoma, T.H., Räsänen, L.A., 1990. Toxic cyanobacteria (blue-green
 215 algae) in finnish fresh and coastal waters. Hydrobiologia 190, 267–275. URL: <https://doi.org/10.1007/bf00008195>,
 216 doi:10.1007/bf00008195.

217 Soares, M.C.S., Lurling, M., Huszar, V.L.M., 2010. Responses of the rotifer *brachionus calyciflorus* to two tropical toxic
 218 cyanobacteria (*cylindrospermopsis raciborskii* and *microcystis aeruginosa*) in pure and mixed diets with green algae.
 219 Journal of Plankton Research 32, 999–1008. URL: <https://doi.org/10.1093/plankt/fbq042>, doi:10.1093/plankt/
 220 fbq042.

221 Staehr, P.A., Birkeland, M.J., 2006. Temperature acclimation of growth, photosynthesis and respiration in two mesophilic
 222 phytoplankton species. Phycologia 45, 648–656. URL: <https://doi.org/10.2216/06-04.1>, doi:10.2216/06-04.1.

223 Sugimoto, K., Negishi, Y., Amano, Y., Machida, M., Imazeki, F., 2015. Roles of dilution rate and nitrogen concen-
 224 tration in competition between the cyanobacterium *microcystis aeruginosa* and the diatom *cyclotella* sp. in eu-
 225 trophic lakes. Journal of Applied Phycology 28, 2255–2263. URL: <https://doi.org/10.1007/s10811-015-0754-7>,
 226 doi:10.1007/s10811-015-0754-7.

227 Suruzzaman, M., Cao, T., Lu, J., Wang, Y., Su, M., Yang, M., 2022. Evaluation of the MIB-producing potential based on
 228 real-time qpcr in drinking water reservoirs. Environmental Research , 112308URL: [https://www.sciencedirect.com/
 229 science/article/pii/S0013935121016091](https://www.sciencedirect.com/science/article/pii/S0013935121016091), doi:10.1016/j.envres.2021.112308.

230 TALBOT, P., 1991. A comparative study and mathematical modeling of temperature, light and growth of three microalgae
 231 potentially useful for wastewater treatment. Water Research 25, 465–472. URL: [https://doi.org/10.1016/0043-
 232 1354\(91\)90083-3](https://doi.org/10.1016/0043-1354(91)90083-3), doi:10.1016/0043-1354(91)90083-3.

233 Tang, E.P.Y., Tremblay, R., Vincent, W.F., 1997. CYANOBACTERIAL DOMINANCE OF POLAR FRESHWATER ECOSYSTEMS:
 234 ARE HIGH-LATITUDE MAT-FORMERS ADAPTED TO LOW TEMPERATURE?1. Journal of Phycology 33, 171–181. URL:
 235 <https://doi.org/10.1111/j.0022-3646.1997.00171.x>, doi:10.1111/j.0022-3646.1997.00171.x.

236 Thomas, M.K., Litchman, E., 2015. Effects of temperature and nitrogen availability on the growth of invasive and na-
 237 tive cyanobacteria. Hydrobiologia 763, 357–369. URL: <https://doi.org/10.1007/s10750-015-2390-2>, doi:10.1007/
 238 s10750-015-2390-2.

239 Tiwari, O., Prasanna, R., Yadav, A., Wattal, D., Singh, P., 2001. Growth potential and biocide tolerance of non-
 240 heterocystous filamentous cyanobacterial isolates from rice fields of uttar pradesh, india. Biology and Fertility of
 241 Soils 34, 291–295. URL: <https://doi.org/10.1007/s003740100402>, doi:10.1007/s003740100402.

242 Torres, C.D.A., Lürling, M., Marinho, M.M., 2016. Assessment of the effects of light availability on growth and com-
 243 petition between strains of *Planktothrix agardhii* and *Microcystis aeruginosa*. Microbial Ecology 71, 802–813.
 244 doi:10.1007/s00248-015-0719-z.

245 Tsai, A.Y., Mukhanov, V., 2021. Response of growth and grazing rate of nanoflagellates on *synechococcus* spp. to experi-

246 mental nutrient enrichment. *Water* 13, 2686. URL: <https://doi.org/10.3390/w13192686>, doi:10.3390/w13192686.

247 Tsujimura, S., Ishikawa, K., Tsukada, H., 2001. Effect of temperature on growth of the cyanobacterium aphanizomenon
248 flos-aquae in lake biwa and lake yogo. *Phycological Research* 49, 275–280. URL: [https://doi.org/10.1111/j.1440-](https://doi.org/10.1111/j.1440-1835.2001.tb00257.x)
249 [1835.2001.tb00257.x](https://doi.org/10.1111/j.1440-1835.2001.tb00257.x), doi:10.1111/j.1440-1835.2001.tb00257.x.

250 Tsujimura, S., Okubo, T., 2003. Development of *Anabaena* blooms in a small reservoir with dense sediment
251 akinete population, with special reference to temperature and irradiance. *Journal of Plankton Research*
252 25, 1059–1067. URL: <http://plankt.oxfordjournals.org/content/25/9/1059.abstract>, doi:10.1093/plankt/25.9.
253 [1059](http://plankt.oxfordjournals.org/content/25/9/1059.full.pdf+html), arXiv:<http://plankt.oxfordjournals.org/content/25/9/1059.full.pdf+html>.

254 Van Der Ploeg, M., Dennis, M., De Regt, M., 1995. Biology of *Oscillatoria chalybea*, a 2-methylisoborneol producing blue-
255 green alga of Mississippi Catfish Ponds. *Water Science and Technology* 31, 173–180. URL: [https://www.sciencedirect.](https://www.sciencedirect.com/science/article/pii/027312239500473Z)
256 [com/science/article/pii/027312239500473Z](https://www.sciencedirect.com/science/article/pii/027312239500473Z), doi:10.1016/0273-1223(95)00473-Z.

257 Venter, A., van Vuuren, S.J., Pieterse, A., 2003. *Oscillatoria simplicissima* : an autecological study. *Wa-*
258 *ter SA* 29, 105–112. URL: <https://journals.co.za/doi/abs/10.10520/EJC116052>, doi:10.10520/EJC116052,
259 [arXiv:https://journals.co.za/doi/pdf/10.10520/EJC116052](https://journals.co.za/doi/pdf/10.10520/EJC116052).

260 Vincent, W., Silvester, W., 1979. Growth of blue-green algae in the manukau (new zealand) oxidation ponds—
261 II. experimental studies on algal interaction. *Water Research* 13, 717–723. URL: [https://doi.org/10.1016/0043-](https://doi.org/10.1016/0043-1354(79)90235-5)
262 [1354%2879%2990235-5](https://doi.org/10.1016/0043-1354(79)90235-5), doi:10.1016/0043-1354(79)90235-5.

263 Wang, W., Song, X.K., Ru, S.G., 2007. Studies on the factors affecting the growth and hemolytic activity of *Anabaena*
264 *variabilis*. *Journal of Applied Phycology* 19, 365–371. URL: <https://doi.org/10.1007/s10811-006-9143-6>, doi:10.
265 [1007/s10811-006-9143-6](https://doi.org/10.1007/s10811-006-9143-6).

266 Wang, Z., Li, R., 2015. Effects of light and temperature on the odor production of 2-methylisoborneol-producing *Pseu-*
267 *danabaena* sp. and geosmin-producing *Anabaena ucrainica* (cyanobacteria). *Biochemical Systematics and Ecology*
268 58, 219–226. URL: <http://www.sciencedirect.com/science/article/pii/S0305197814003378>, doi:10.1016/j.bse.
269 [2014.12.013](http://www.sciencedirect.com/science/article/pii/S0305197814003378).

270 Watanabe, M.F., Oishi, S., 1985. Effects of environmental factors on toxicity of a cyanobacterium (*Microcystis aerugi-*
271 *nosa*) under culture conditions. *Applied and Environmental Microbiology* 49, 1342–1344. URL: [https://doi.org/10.](https://doi.org/10.1128/aem.49.5.1342-1344.1985)
272 [1128/aem.49.5.1342-1344.1985](https://doi.org/10.1128/aem.49.5.1342-1344.1985), doi:10.1128/aem.49.5.1342-1344.1985.

273 van der Westhuizen, A.J., Eloff, J.N., 1985. Effect of temperature and light on the toxicity and growth of the blue-green
274 alga microcystis aeruginosa (UV-006). *Planta* 163, 55–59. URL: <https://doi.org/10.1007/bf00395897>, doi:10.1007/
275 [bf00395897](https://doi.org/10.1007/bf00395897).

276 Wiedner, C., Visser, P.M., Fastner, J., Metcalf, J.S., Codd, G.A., Mur, L.R., 2003. Effects of light on the microcystin content
277 of *Microcystis* strain PCC 7806. *Applied and Environmental Microbiology* 69, 1475–1481. URL: [https://doi.org/10.](https://doi.org/10.1128/aem.69.3.1475-1481.2003)
278 [1128/aem.69.3.1475-1481.2003](https://doi.org/10.1128/aem.69.3.1475-1481.2003), doi:10.1128/aem.69.3.1475-1481.2003.

279 Willis, A., Chuang, A.W., Woodhouse, J.N., Neilan, B.A., Burford, M.A., 2016. Intraspecific variation in growth, mor-
280 phology and toxin quotas for the cyanobacterium, *cylindrospermopsis raciborskii*. *Toxicon* 119, 307–310. URL:

281 <https://doi.org/10.1016/j.toxicon.2016.07.005>, doi:10.1016/j.toxicon.2016.07.005.

282 Wu, X., Wu, Z., Song, L., 2011. Phenotype and temperature affect the affinity for dissolved inorganic carbon in
283 a cyanobacterium microcystis. *Hydrobiologia* 675, 175–186. URL: <https://doi.org/10.1007/s10750-011-0815-0>,
284 doi:10.1007/s10750-011-0815-0.

285 Xiao, M., Hamilton, D.P., O'Brien, K.R., Adams, M.P., Willis, A., Burford, M.A., 2020. Are laboratory growth rate experi-
286 ments relevant to explaining bloom-forming cyanobacteria distributions at global scale? *Harmful Algae* 92, 101732.
287 URL: <https://doi.org/10.1016/j.hal.2019.101732>, doi:10.1016/j.hal.2019.101732.

288 Xiao, M., Willis, A., Burford, M.A., 2017. Differences in cyanobacterial strain responses to light and temperature reflect
289 species plasticity. *Harmful Algae* 62, 84–93. URL: <https://doi.org/10.1016/j.hal.2016.12.008>, doi:10.1016/j.hal.
290 2016.12.008.

291 Zhang, T., Li, L., Song, L., Chen, W., 2009. Effects of temperature and light on the growth and geosmin production of
292 *Lyngbya kuetzingii* (cyanophyta). *Journal of Applied Phycology* 21, 279–285. URL: [http://dx.doi.org/10.1007/s10811-](http://dx.doi.org/10.1007/s10811-008-9363-z)
293 [008-9363-z](http://dx.doi.org/10.1007/s10811-008-9363-z), doi:10.1007/s10811-008-9363-z.

294 Zhang, T., Zheng, L., Li, L., Song, L., 2016. 2-methylisoborneol production characteristics of *Pseudanabaena* sp. FACHB
295 1277 isolated from Xionghe Reservoir, China. *Journal of Applied Phycology* , 1–10URL: [http://dx.doi.org/10.1007/](http://dx.doi.org/10.1007/s10811-016-0864-x)
296 [s10811-016-0864-x](http://dx.doi.org/10.1007/s10811-016-0864-x), doi:10.1007/s10811-016-0864-x.